Supplement 1: Summary of *in situ* investigations of soil water depth-profiles, bulk soil water isotopes, xylem water isotopes and transpired water isotopes (arranged by these four groups, within groups chronological order)

Author	Environment / Location	Measured compartment	Method	Findings/ Recommendations Advantages (+) /Disadvantages (-)
(Herbstritt et al., 2012)	Laboratory	Liquid water Soil column	<ul> <li>Material: gas permeable membrane (PP)</li> <li>Soil type: sand</li> <li>Push-through system, flow rate 200 ml/min with excess line, equilibrated with pure N<sub>2</sub></li> <li>Calibration: <ul> <li>liquid water standards injected</li> <li>compared to vapor standards</li> <li>correction for water vapor concentration</li> <li>determination of fractionation factors</li> <li>application in soil column experiment</li> </ul> </li> </ul>	<ul> <li>Fit equilibrium fractionation factors for each system (membrane effects)</li> <li>Vapor concentration and temperature are highly correlated</li> <li>Matric effects might have caused deviation of fractionation factors, not membrane effects</li> <li>Temporal resolution of less than 1 min. for liquid water samples</li> </ul>
(Soderberg et al., 2012)	Semiarid, Kenya	Soil water isotope depth-profiles	<ul> <li>Material: gas permeable membrane (Teflon) with open ending, packed with glass wool</li> <li>Soil type: sandy loam</li> <li>Pull-only system with additional dilution of 400 ml/min, diluted with ambient air</li> <li>Calibration/Validation: No information provided</li> </ul>	<ul> <li>Include soil water potential into Craig-Gordon model for soils</li> <li>Hncludes modelling of results</li> <li>Glass wool might fractionate</li> <li>Condensation issue not addressed</li> <li>Dilution with non-dried ambient air</li> </ul>
(Volkmann and Weiler, 2014)	Humid (southwest Germany)	Soil water isotope depth-profiles Evolution of evaporation front	<ul> <li>Material: gas permeable membrane (PE) with mixing chamber</li> <li>Soil type: clayey silt</li> <li>Both push-trough (flow rate not specified) and pull-only (30-35 ml/min) systems used, dilution with pure N2</li> <li>Calibration: <ul> <li>boxes filled with soil and equipped with gas probes, water content, EC, temperature, soil water potential sensors</li> <li>isotope standards added to water content of 20 vol%</li> <li>linear two-point calibration &amp; quality control standard</li> <li>correction for vapor-concentration dependence</li> <li>liquid-vapor equilibrium after Majoube (1971)</li> <li>normalization to VSMOW and drift correction</li> </ul> </li> </ul>	<ul> <li>First successful field study and comprehensive investigation of <i>in situ</i> soil water isotopes</li> <li>Very flexible system</li> <li>Both stationary and mobile use</li> <li>Both push-through and pull-only methods tested</li> <li>Control of condensation via dilution directly in probe</li> <li>Complex system</li> <li>Expensive probes</li> </ul>
(Rothfuss et al., 2015, 2013)	Laboratory	Soil water isotopes	<ul> <li>Material: gas permeable membrane (PP)</li> <li>Soil type: fine sand</li> <li>Push-trough system with excess tube; dry synthetic air flow of 25 ml/min at inlet, dilution with dry synthetic air after passage through soil</li> <li>Calibration: <ul> <li>three standards prepared, added to dried soil and measured over a range of temperatures (8°C - 24°C)</li> <li>constant water vapor concentration via dilution with dry synthetic air</li> <li>drift correction and check for material changes of the soil gas probe</li> </ul> </li> <li>Validation: comparison to theoretically calculated vapor values</li> </ul>	<ul> <li>Potential isotope-effects introduced via gas permeable probe (difference of theoretical and observed values for δ<sup>2</sup>H<sub>vap</sub>)</li> <li>Uncertainties when using CG-model</li> <li>Gas-permeable membranes can resolve rapid changes of isotope values</li> <li>Linear correction for non-reached isotopic equilibrium</li> <li>Water vapor concentration kept constant</li> <li>Checked for equilibrium conditions</li> <li>Test limited to laboratory conditions</li> <li>Isotopic equilibrium not reached</li> </ul>

(Gangi et al., 2015)	Laboratory	Soil water and carbon dioxide δ <sup>18</sup> O	<ul> <li>Material: gas permeable membrane (PP)</li> <li>Soil type: fine sand</li> <li>Push-trough system with excess tube; dry synthetic air flow of 85 ml/min at inlet, dilution with dry synthetic air after passage through soil</li> <li>Validation: comparison to theoretically calculated and modelled values</li> </ul>	<ul> <li>No fractionation due to tubing for δ18O</li> <li>Combined measurements of oxygen isotopologues in soils and atmospheric carbon dioxide</li> <li>+ modelling involved (MuSICA)</li> <li>- No field testing</li> </ul>
(Gaj et al., 2016)	Semi-arid (Namibia)	Soil water isotope depth-profiles Evolution of evaporation front	<ul> <li>Material: gas permeable membrane (PP)</li> <li>Soil type: medium sand</li> <li>Pull-only at 95 to 110 ml/min, pre-flushing of probes using dry air; system changeable to push-through</li> <li>Calibration: <ul> <li>four standards prepared in oven-dried soil material; 5 ml of standard solution added</li> <li>drift and memory correction, quality control</li> <li>conversion of the corrected vapor to respective liquid water isotopic compositions</li> </ul> </li> <li>Validation: depth profiles using cryogenic vacuum extraction</li> </ul>	<ul> <li>First successful <i>in situ</i> study in semi-arid environment</li> <li>Recommendations: <ul> <li>Organic contamination should be investigated further</li> <li>Increase probe length under dry conditions</li> <li>Use heating/dilution to avoid condensation</li> <li>Further development of calibration strategies for clay-rich soils.</li> <li>calibration in soil media</li> <li>water vapor concentration controlled by sampling time</li> <li>calculated influence radius for pull-only method and air volumes sampled</li> <li>Not corrected for water vapor concentration</li> <li>Strong deviations in upper soil layer</li> </ul> </li> </ul>
(Pratt et al., 2016)	Humid (Canada)	Waste rock mining site unsaturated zone isotopes	<ul> <li>Material: HDPE tubes with 50-mm stainless steel mesh filter</li> <li>Soil type: coarse textured sand tailings dyke</li> <li>Pull-only, 30 ml/min, no dilution or flushing</li> <li>Calibration: <ul> <li>10ml of liquid water standards added to Ziploc bag</li> <li>correction for water vapor concentration</li> <li>conversion of the corrected vapor to respective liquid water isotopic compositions</li> </ul> </li> <li>Validation: comparison to bag equilibration method</li> </ul>	<ul> <li>Direct <i>in situ</i> measurement of natural, water stable isotope profiles thick unsaturated mine waste is challenging</li> <li>Bag equilibration as alternative</li> <li>General agreement to samples collected in equilibration bags for some depths</li> <li>Strong deviations in most depths</li> </ul>
(Oerter et al., 2019, 2017; Oerter and Bowen, 2017, 2019)	Desert/Arid (Utah, United States)	Soil water isotope depth-profiles	<ul> <li>Material: gas permeable membrane (PP)</li> <li>Soil type: coarse textured sand tailings dyke</li> <li>Push-trough system with excess tube; dry N<sub>2</sub> for flushing and dilution; flow of 60 ml/min; dilution flow of 20 ml/min</li> <li>Calibration: <ul> <li>oven dried (105°C for &gt;12 h) soil material at 7%, 18%, 26% GWC (g water g<sup>-1</sup> soil), n = 18 calibration standards (3 soil depth groups (7% to 11% clay content) × 3 GWC contents × 2 water types)</li> <li>multi-linear regression for one-step correction of water isotope data derived from standards, variables used: clay content, water content, water vapor isotope value</li> <li>CO<sub>2</sub> spectral interference correction</li> </ul> </li> <li>Validation: depth profiles using cryogenic vacuum extraction and bag equilibration</li> </ul>	<ul> <li>Throughput rates greater than 7h<sup>-1</sup></li> <li>Accuracy of the method to be equivalent to direct headspace equilibration and vacuum extraction techniques</li> <li>measurement duration of ~5 min is typically sufficient to achieve &gt;2 min of stable H<sub>2</sub>O and δ<sup>2</sup>H and δ<sup>18</sup>O measurements</li> <li>Most complete approach in terms of calibration and setup</li> <li>water vapor concentration kept constant (20.000 ppm)</li> <li>calibration carried out in same media as measured (=soil)</li> <li>one-step calibration with different soil types and water contents; calibration can easily be extended to more variables</li> </ul>

				- Effect of changing environmental conditions (temperature) not addressed, issue of condensation not addressed at all
(Kühnhammer et al., 2019)	Laboratory	Water isotopes across soil column and in plant transpiration	<ul> <li>Material: gas permeable membrane (PP) and throughflow plant chamber</li> <li>Soil type: silt loam</li> <li>Plant species: <i>Centaurea jacea</i> (herbaceous)</li> <li>Push-trough system with excess tube; dry synthetic air flow of 50 ml/min at inlet, dilution with dry synthetic air</li> <li>Calibration: <ul> <li>two soil standards with gas permeable membranes integrated into automated system</li> <li>calibration equation developed by Rothfuss et al (2013), drift correction, correction for effect of wvmr</li> </ul> </li> <li>Validation: <ul> <li>comparison of calculated liquid values with water added to soil column</li> <li>agreement between soil and transpiration isotope values in dual isotope space</li> <li>comparison root water uptake profiles and changes in soil water content</li> </ul> </li> </ul>	<ul> <li>Data set of 48 days monitoring isotopic changes in soil and plant water uptake</li> <li>+ error propagation to estimate realistic value for measurement precision</li> <li>+ combined continuous <i>in situ</i> measurement across soil profile and in plant transpiration</li> <li>+ check for isotopic steady-state in plant transpiration</li> <li>+ root water uptake modelling including additional parameters</li> <li>- No check for isotopic equilibrium in soil measurements</li> <li>- Not easily transferable to the field</li> </ul>
Dubbert et al., 2013	Semi-arid (Portugal)	Evaporation bulk water isotopes	<ul> <li>Material: Custom build soil chamber</li> <li>Soil type: loamy sand</li> <li>Open gas exchange system following the design of Pape et al. (2009)</li> <li>Calibration: <ul> <li>liquid injection of 3 standards</li> <li>water vapor concentration dependency from 5000-30000 ppmv considered</li> </ul> </li> </ul>	<ul> <li>First data set observing soil evaporation <i>in situ</i> under field conditions</li> <li>Sensitivity analysis of the Craig and Gordon model for input parameters</li> <li>Different conditions monitored (bare soil, root ingrowth, vegetated soil)</li> <li>High impact of vegetation cover and root ingrowth on isotopic signature of soil evaporation</li> </ul>
Quade et al. 2018	Laboratory	Evaporation bulk water isotopes	<ul> <li>Material: Custom build soil chamber</li> <li>Soil type: silty loam</li> <li>Keeling plot approach</li> <li>Calibration: <ul> <li>Vapor equilibration standards in the same soil as the experimental column</li> </ul> </li> </ul>	<ul> <li>Evaluating temporal dynamics of the kinetic fractionation factor</li> <li>αK values within the range reported in the literature</li> <li>prevalence of turbulent water vapor transport under water-saturated soil conditions and at soil water content significantly lower than saturation</li> </ul>
(Volkmann et al., 2016)	Humid (Southern Germany	Xylem water isotopes	<ul> <li>Material: gas permeable membrane (PE) with mixing chamber</li> <li>Plant species: deciduous maple trees (<i>Acer campestre L</i>.; 9m tall, 0.2m in diameter at breast height and 30m<sup>2</sup> of crown projected area)</li> <li>Mixed system; dry N<sub>2</sub> provided in throughflow line, set at the flow rate induced by laser spectrometer (30-35 ml/min); dilution with N<sub>2</sub> directly in probe; pre-flushing of system for 120s</li> <li>Calibration: <ul> <li>three standards, headspace measured; conversion of xylem vapor values using Majoube equation and measured temperatures in tree</li> </ul> </li> <li>Validation:</li> </ul>	<ul> <li>First study of <i>in situ</i> measured xylem water isotopes</li> <li><i>In situ</i> monitoring of xylem water isotopes is feasible, but complicated</li> <li>Good precision and repeatability, but accuracy needs to be improved</li> <li>Longer-term study needed</li> <li>Issue of organic contamination needs to be addressed</li> <li>+ Condensation avoided via dilution directly in probe</li> <li>No check if equilibration conditions were reached in probe</li> <li>Partially large discrepancies between extracted values and in situ measured data</li> <li>- Complicated</li> </ul>

			<ul> <li>comparison with vacuum-extracted xylem cores measured with IRMS</li> </ul>	
(Marshall et al.)	Greenhouse	Xylem water isotopes	<ul> <li>Material: hole drilled through the stem of a tree, Swagelok connections on both sides (airtight); flush with acetone to avoid pitch and resin production</li> <li>Plant species: Pine trees (<i>Pinus sylvestris</i> L. and <i>Pinus pinea</i> L.); first experiment: cut-stem tree; second experiment: intact-root experiment</li> <li>First experiment: pull-only with ~35-40 ml/min and flush of system manually when needed; second experiment: push-through with 80 ml/min of dry air (diving air), 5min pre-flush and 15 min measurement time</li> <li>Calibration: <ul> <li>four standards, headspace measured; conversion of xylem vapor values using Majoube equation and measured temperatures in tree</li> </ul> </li> <li>Validation: <ul> <li>comparison to source water isotope values</li> </ul> </li> </ul>	<ul> <li>Novel approach for measuring xylem water isotopes</li> <li>Method requires testing for different tree species and under different environmental conditions</li> <li><i>In situ</i> monitoring of xylem water isotopes of natural abundances is possible</li> <li>No gas-permeable membranes are required</li> <li>No influence of organic contamination observed</li> <li>fundamental aspects of <i>in situ</i> studies thoroughly evaluated by modelling</li> <li>Systems needs to be simplified</li> <li>possibility of liquid water reaching water vapor analyser if tree shows strong defence mechanism or 'backflow' of water occurs</li> </ul>
Wang et al., 2013	Great Plains, Oklahoma, USA	Evaporation bulk water isotopes Transpiration water isotopes Evapotranspiration water isotopes	<ul> <li>Material: modified commercial soil chamber, commercial plant chamber, custom build ET chamber</li> <li>Soil type: not provided</li> <li>Plant species: <i>Bromus arvensis L., Vicia sativa L., Solanum</i> <i>carolinense L., Euphorbia dentata Michx., Tridens flavus (L.)</i> <i>Hitchc.</i></li> <li>Keeling plot approach</li> <li>Calibration: Liquid injections</li> </ul>	<ul> <li>Testing chamber based bare soil observations vs modelled by Craig and Gordon model and impact on ET partitioning</li> <li>Difference between approaches on T/ET</li> <li>Demonstrate necessity for uniform partitioning approach</li> </ul>
(Wang et al., 2012)	Laboratory, arid (Kenia)	Transpiration water isotopes	<ul> <li>Material: custom build transpiration chamber, OA-ICOS</li> <li>Plant species: <i>Spathiphyllum spp, Acacia spec</i>.</li> <li>Open gas exchange system</li> <li>Calibration: <ul> <li>verification of set up using a dew point generator</li> <li>liquid injections to calibrate the laser spectrometer</li> </ul> </li> </ul>	<ul> <li>First publication of in-situ transpiration measurements</li> <li>+ Observation of transit from non-steady state to isotopic steady state</li> <li>- Limited data set</li> <li>- Complex set up</li> </ul>
(Simonin et al., 2013)	laboratory	Transpiration water isotopes	<ul> <li>Material: fully automated leaf gas exchange system (MPH1000, Licor 7600); Laser spectrometry</li> <li>Plant species: Citrus, Tobacco</li> <li>Open gas exchange system</li> <li>Calibration: <ul> <li>Liquid water standards injected spanning the observed isotopic range</li> <li>Concentration dependencies 3000-24000 ppmv</li> </ul> </li> </ul>	<ul> <li>Exposed leaves to changes in environmental conditions</li> <li>Non-steady state effects dominate transpiration of both species</li> <li>Even when leaves are already physiologically transpiring in steady state</li> <li>Evaluated the rate of change in leaf water, determined leaf water turn-over time</li> </ul>
(Dubbert et al., 2014)	Semi-arid (central Portugal)	Transpiration water isotopes	<ul> <li>Material: Custom build branch chamber, Laser spectrometer</li> <li>Plant species: cork oak (<i>Quercus suber L.</i>)</li> <li>Open gas exchange system</li> <li>Calibration: <ul> <li>liquid injection of 3 standards</li> <li>concentration dependency from 5000-30000 ppmv</li> </ul> </li> </ul>	<ul> <li>Diurnal courses of isotopic signatures of transpiration during different seasons</li> <li>Dominance of non-steady state effects</li> <li>Sensitivity analysis of the Craig and Gordon/Dongmann based model</li> </ul>
(Song et al., 2015)	Laboratory	Transpiration water isotopes	• Material: Fully automated leaf gas exchange system (Licor 7600), Laser spectrometer	• Adapting existing non-steady state leaf water model to cuvettes, where delta atm is influenced by delta T

	Plant species: Cotton	
	• Calibration:	
	- liquid injection, following Simonin et al., 2013	