

1 **Carbon and Nitrogen Dynamics and Greenhouse Gas Emissions in Constructed**
2 **Wetlands Treating Wastewaters: A Review**

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34 **Abstract**

35 The removal efficiency of carbon (C) and nitrogen (N) in constructed wetlands (CWs) is very
36 inconsistent and frequently does not reveal whether the removal processes are due to physical
37 attenuation or whether the different species have been transformed to other reactive forms. This
38 paper aims to address this knowledge gap by reviewing the biogeochemical dynamics and fate of
39 C and N in CWs and their potential impact on the environment, and by presenting novel ways in
40 which these knowledge gaps may be eliminated. Nutrient removals in CWs vary with the type of
41 CW, vegetation, climate, season, geographical region and management practices. Horizontal flow
42 CWs tend to have good nitrate (NO_3^-) removal, as they provide good conditions for
43 denitrification, but cannot remove ammonium (NH_4^+) due to limited ability to nitrify NH_4^+ .
44 Vertical flow CWs have good NH_4^+ removals, but their denitrification ability is low. Surface flow
45 CWs decrease nitrous oxide (N_2O) emissions but increase methane (CH_4) emissions; subsurface
46 flow CWs increase N_2O and carbon dioxide (CO_2) emissions, but decrease CH_4 emissions. Mixed
47 species of vegetation perform better than monocultures in increasing C and N removal and
48 decreasing GHG emissions, but empirical evidence is still scarce. Lower hydraulic loadings with
49 higher hydraulic retention times enhance nutrient removal, but more empirical evidence is
50 required to determine an optimum design. A conceptual model highlighting the current state of
51 knowledge is presented and experimental work that should be undertaken to address knowledge
52 gaps across CW, vegetation and wastewater types, hydraulic loading rates and regimes, and
53 retention times, is suggested. We recommend that further research on process-based C and N
54 removal and on the balancing of end products into reactive and benign forms is critical to the
55 assessment of the environmental performance of CWs. While nutrient cycles have been addressed
56 in previous books and review papers, we strongly suggest that *in situ* transformation and fate
57 of the transformation products with regard to pollution swapping requires further detailed
58 examination.

59
60 Keywords: Carbon, nitrogen, constructed wetlands, pollution swapping, nitrous oxide, methane

61
62 1. Introduction

63 Increasing anthropogenic loading of reactive nitrogen (Nr; all forms of nitrogen except di-
64 nitrogen gas, N_2) along the nitrogen (N) cascade in the environment raises many critical concerns
65 for human health, drinking water quality (Gray, 2008), coastal and marine water degradation, as
66 well as algal blooms and hypoxia (Conley et al., 2009; Rabalais et al., 2010). While there are

67 excellent reviews and books available on the biogeochemistry of wetlands in general (Reddy and
68 Delaune, 2008; Kadlec and Wallace, 2009), less is known about constructed wetlands (CWs),
69 which provide much more controlled conditions (Vymazal and Kröpfelova, 2010). Constructed
70 wetlands (CWs) are artificial sinks for Nr (Galloway et al, 2003; Tanner et al., 2005), and have
71 been successfully used for treating domestic sewage, urban runoff and storm water, industrial and
72 agricultural wastewater, and leachate. Although CWs have a proven potential for organic carbon
73 (C) and N removal, with few exceptions, studies have not adequately delineated removal
74 pathways. This has meant that reported removal efficiencies have been variable (Seitzinger et al.,
75 2002), and appropriate design and management strategies have not been adopted.

76

77 Constructed wetlands are complex bioreactors that facilitate a number of physical, chemical and
78 biological processes, but are frequently evaluated as a 'black box' in terms of process
79 understanding (Langergraber, 2008). Many investigations target single contaminant remediation
80 and disregard the reality of mixed contaminants entering and leaving CWs. They do not consider
81 the dynamics of "pollution swapping" (the increase in one pollutant as a result of a measure
82 introduced to reduce a different pollutant) driven by transformational processes within and
83 around the system. This means that potential negative impacts that CWs may have on the
84 environment, such as greenhouse gas (GHG) emissions (IPCC, 2013; Clair et al., 2002; Mander
85 et al., 2008; Mitsch and Gosselink, 2000) or enhancement of pollution swapping (Reay, 2004),
86 are not accounted for in analyses. There are many pathways by which the removed N can
87 contribute to water and air pollution: accumulation and adsorption in soils, leaching of nitrate
88 (NO_3^-) and ammonium (NH_4^+) to groundwater, emissions of nitrous oxide (N_2O) and ammonia
89 (NH_3^+) to the atmosphere, and/or conversion to N_2 gas. Constructed wetlands significantly
90 contribute to atmospheric N_2O emissions either directly to the atmosphere from the surface of the
91 wetland (IPCC, 2013; Søvik et al., 2006; Ström et al., 2007; Elberling et al., 2011) or indirectly
92 *via* dissolved N_2O in the effluent or groundwater upon discharge to surface waters. The IPCC
93 (2013) has recognised the significance of indirect N_2O emissions from CW effluent that is
94 discharged to aquatic environments, and estimate emission factors (EF) ranging from 0.0005 to
95 0.25. Production and reduction processes of N_2O in the environment are not yet fully understood.

96

97 Constructed wetlands receive organic C from the influent wastewater and from fixation by the
98 photosynthetic hydrophytes, which are incorporated into soil as organic C. Soil organic C
99 undergoes the biogeochemical processes that regulate C accretion in soil and microbial
100 respiration, producing carbon dioxide (CO_2). Anaerobic mineralization of organic C by

101 methanogenic archaea can produce methane (CH₄) (Laanbroek, 2010; Ström et al., 2007; Søvik et
102 al., 2006; Pangala et al., 2010). Constructed wetlands can also contribute to the dissolved organic
103 carbon (DOC) load transfer to ground and surface waters, which may produce and exchange
104 substantial amounts of CO₂ and CH₄ with the atmosphere (Clair et al., 2002; Elberling et al.,
105 2011). Therefore, CWs can diminish the environmental benefits of wastewater treatment. The
106 dynamics of dissolved N₂O, CO₂ and CH₄ in CWs is a key knowledge gap in global greenhouse
107 gas budgets.

108
109 Surface emissions of GHG from CWs have been commonly measured by the closed chamber
110 method (Johansson et al., 2003, 2004; Mander et al., 2005, 2008), but have rarely been measured
111 by ebullition and diffusion methods (Søvik et al., 2006). The measured rates have shown high
112 spatial, temporal and diurnal variations due to the change in biogeochemistry of C and N and
113 plant-microbe-soil interaction over time and space. Surface emissions cannot explain the kinetics
114 of production and consumption rates of GHG, which we need to know in order to adopt better
115 management practices to mitigate emissions. In addition, subsurface export of dissolved nutrients
116 and GHG, an important pathway of nutrient loss (Riya et al., 2010), is frequently ignored. Mass
117 balance analysis of the different components of the N cycle and kinetics of their transformation
118 processes occurring within the treatment cells using the isotope-tracing ¹⁵N technique can provide
119 mechanistic information for N transformation products (Lee et al., 2009; O’Luanaigh et al., 2010)
120 and may be used to start to answer such questions. Similarly, ¹⁴C application and measurement of
121 C species (e.g. CO₂, CH₄, and DOC) may elucidate the C mineralization and CO₂ and CH₄
122 production and consumption. Used in combination, these methods may provide a comparative
123 analysis of the rates of C and N transformation processes and role of these processes in delivering
124 NO₃⁻, NH₄⁺ and DOC to ground/surface waters and N₂O, CO₂ and CH₄ to the atmosphere.

125
126 Past reviews on CWs, though very limited, summarise the performance of different types of CWs
127 on C and N removal (Vymazal, 2007) and surface emissions of GHG (Mander et al., 2014), but
128 have not discussed the mechanisms of nutrient removal and the fate of the nutrients delivered and
129 removed to and from CWs. Therefore, the objectives of this review are (i) to understand the
130 biogeochemical dynamics of C and N in CWs to (ii) better understand the fate of various C and N
131 species in a holistic manner, in addition to the conventional influent/effluent balance for nutrient
132 removal (iii) identify the research gaps that need to be addressed to optimise nutrient removal and
133 mitigate GHG emissions, and (iv) discuss emerging measurement techniques that may give
134 insights into the production and reduction of GHG.

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2. Removal Efficiency, Hydraulic Loading and Retention Time

In CWs, the efficiency of C and N removal is generally limited and highly variable over CW types, plant types, seasons, climatic regions and management practices. On average, it appears that 50 and 56% of the influent total nitrogen (TN) and total organic carbon (TOC), respectively, can be removed, but the removal rates are very inconsistent. Mean (\pm standard error) TN removals, obtained from the literature cited in this paper, ranged from $31.3 \pm 6.3\%$ in surface flow (SF) CWs to $40.4 \pm 4.4\%$ in subsurface flow CWs, whereas TOC removal ranged from $18.8 \pm 9.4\%$ in SF CWs to $56.2 \pm 9.5\%$ in vertical subsurface flow CWs (Table 1 and Table 2). In European systems, for example, typical removals of ammoniacal-N in long-term operation are around 35%, but can be enhanced if some pre-treatment procedures are followed (Verhoeven, and Meuleman, 1999; Luederitz et al., 2001). Generally, TN removal is higher in SF CWs than subsurface flow (SSF) CWs (Table 1), but studies differ. For example, Van der Zaag et al. (2010) showed higher N removal in SF CWs than SSF, but Søvik et al. (2006) and Gui et al. (2007) showed the opposite. In SSF CWs, limited removal can be caused by a reduced environment that enhances NH_4^+ accumulation and limits NH_4^+ oxidation. In SF CWs, denitrification rates can be limited due to lack of NO_3^- . In vertical subsurface flow (VSSF) CWs, aeration can increase NH_4^+ oxidation to NO_3^- , which can be denitrified or converted to NH_4^+ by dissimilatory NO_3^- reduction to NH_4^+ (DNRA).

Plant species are important components of CWs, and affect C and N removals. Optimal species selection for best removal is difficult because some species are efficient in removing one pollutant but not the other (Bachand and Horne, 2000; Bojcevska and Tonderski, 2007; da Motta Marques et al., 2000). In some studies there are no inter-species differences at all (Calheiros et al., 2007). Mixed species perform better than monocultures to remove C and N pollutants because they increase microbial biomass and diversity. Payne et al. (2014a) has discussed the role of plants in nutrient removal. Plants regulate CWs hydrology (evaporation and transpiration) and temperature (insulating CWs from seasonal temperature change, trapping falling and drifting snow and heat loss of wind). Some species can create large surface area for microbial attachment and enhance microbial diversity, but experimental evidence is still scarce.

Soil physicochemical properties are important factors controlling the purification capacity in CWs. Soils with high permeability enhance downward nutrient movement to groundwater. High cation exchange materials in soil enhance NH_4^+ fixation by the soil matrix. Microbial activities

169 and growth depend on substrate C quality and C:N ratios, which affect nutrient removal. Better
170 growth of heterotrophic microorganisms is a function of the wastewater C:N (Makino et al.,
171 2003). Higher C:N ratios can enhance denitrification by providing electron donors for denitrifiers,
172 but the opposite can increase nitrification. High C:N ratios can also encourage DNRA over
173 denitrification. Yan et al. (2012) measured a high TN removal but low TOC removals at a C:N
174 ratio 2.5:1, which indicates that removal of one parameter might lead to a problem with a
175 different one. The uncertainty in the conditions for achievement of optimum removal suggests
176 that the rates of C and N transformations and the fate of the removed nutrients within CWs
177 should be investigated. However, to our knowledge, no study has provided a holistic evaluation
178 of C and N attenuation and transformation.

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180 The removal of pollutants in CWs depends on hydraulic loading rates (HLR) and hydraulic
181 retention time (HRT) (Toet et al., 2005). The HLR and HRT are considered to be significant
182 design parameters determining the nutrient removal efficiencies (Weerakoon et al., 2013). Longer
183 residence time of wastewater in CWs increases the removal of C and N (Wang et al., 2014) by
184 increasing sedimentation and duration of contact between nutrients and the CWs. The effects of
185 HLR and HRT can vary with the nature of the use of CWs e.g., whether they are used for treating
186 single or mixed pollutants. To reduce N_r delivery to the receiving waters or to the atmosphere,
187 CWs need to be optimally designed with respect to HLR and HRT.

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189 Fluctuating hydraulic loading influences all biotic and abiotic processes in CWs. For example, if
190 the groundwater table is lowered through changes in hydraulic loading, soil aeration can increase
191 or decrease. Ammonification and nitrification rates increase with increased soil aeration and this
192 enhances C utilization by bacteria and, therefore, can stimulate the removal of C and N.
193 Investigation into the effects of fluctuating hydraulic loadings (“hydraulic pulsing”) on C and N
194 removals and their transformation products will provide information about the fate of the added
195 nutrients in terms of their environmental benefits and/or pollution swapping potential. For
196 example, if the dominant product is N₂, the system will be relatively benign in terms of its impact
197 on the environment, but if it is NH₄⁺, it can be fixed in the soils or transported to ground and
198 surface waters connected to CWs if the cation exchange sites become saturated. Several authors
199 have used a wide range of HLRs and HRTs to measure nutrient removal efficiency, but
200 experimental evidence linking HLR and HRT to removal efficiency is scarce (Toet et al., 2005).
201 Luo et al. (2005) reported that low HLR results in incomplete denitrification, whereas Zhang et
202 al. (2006) argued that low HLR increases NH₄⁺ and chemical oxygen demand oxidation. The way

203 in which the performance of a CW is assessed can lead to different conclusions regarding the
204 removal of Nr. For future studies, evaluation of CWs in a holistic manner, which includes
205 pollution swapping at different HLRs and HRTs, is important, particularly within the context of
206 the changing hydrologic cycle in a changing climate. In addition, local legislative targets should
207 be considered and weighting factors (e.g. the relative importance of, say, GHG over water quality
208 targets) should be developed to evaluate the overall performance of CWs. In addition to the
209 estimation of nutrient removal rates, investigation of the effect of HLR and HRT on the different
210 forms of nutrients in the final effluent and their fate in the natural environment may help
211 elucidate the pollution swapping potential of CWs.

213 3. Accumulation of C and N in CWs Soils

214 The soil in CWs is a major sink for C and N (Mustafa and Scholz, 2011). However, although data
215 on the influent and effluent N concentrations are available, data on N accumulation (dissolved
216 organic nitrogen (DON), TN, NH_4^+ or NO_3^- -N) within the soil profile of various CWs are scarce.
217 The wide range of N accumulation reported in the literature (e.g. 30-40%, Shamir et al., 2001;
218 39%, Harrington et al., 2007; 9%, Mander et al., 2008; 2.5%, Obarska-Pempkowiak and
219 Gajewska, 2003) may be due to the variations in CW types and management strategies. The
220 accumulated species of N are reactive unless they have been transformed to N_2 by
221 biogeochemical processes. However, there is a dearth of information on the extent of Nr
222 accumulation in soils and discharge to surface waters and air (Shamir et al., 2001). Accumulated
223 organic N could be mineralised to NH_4^+ and NO_3^- , depending on the physico-chemical properties
224 of soil. The Nr could be assimilated by plants and microbes, which are recycled in a soil-plant-
225 soil continuum. Nitrogen spiralling occurs from NH_4^+ to organic N and back to NH_4^+ within the
226 CW (O'Lunaigh et al., 2010). Typically, N accumulation has been found to decrease with soil
227 depth (Shamir et al., 2001). In terms of the conventional input-output balance, these are
228 considered as removed N, but may, in fact, remain in such a biogeochemically active system. In
229 addition to N, organic C accumulation occurs in CW soils (Nguyen, 2000). As such, soils of CWs
230 represent organic C and Nr-rich systems, where the products of the continuously occurring
231 biogeochemical processes can be transported to fresh waters and to the atmosphere. Estimation of
232 the rates of nutrient accumulation in soils in various types of CWs under different management
233 systems is important. The stability of the accumulated C and N under changing climatic scenarios
234 also needs to be addressed to consider the long-term sustainability of CWs.

236 4. C and N Dynamics and Greenhouse Gas Emissions

237 Increased nutrients input to the CWs increases the productivity of wetland ecosystems and the
238 production of GHG. As CWs are designed to remove pollutants in an anaerobic/suboxic
239 environment, they change the C and N biogeochemistry and contribute significantly to CH₄ and
240 N₂O emissions (Johansson et al., 2002, 2003; Mander et al., 2005, 2008; Stadmark and
241 Leonardson, 2005; Liikanen et al., 2006). Sövic et al. (2006) measured N₂O, CH₄ and CO₂
242 emissions in various CWs in different European countries, and suggested that the potential
243 atmospheric impacts of CWs should be examined as their development is increasing globally.
244 Management of CWs must consider the negative climatic aspects of increased emissions of GHG
245 in addition to their primary functions (Ström et al., 2007). Therefore, estimation of the
246 contribution of CWs to global warming is required. In this regard, measurement of spatial and
247 temporal variations (seasonal and diurnal) of GHG emissions is necessary to accurately estimate
248 CW-derived GHG emissions. A holistic assessment of ecologically engineered systems has been
249 outlined by Healy et al. (2011, 2014) and developed further by Fenton et al. (2014). Such
250 assessments can be applied in evaluating nutrient dynamics in CWs. Moreover, plant mediated
251 GHG emissions could be an important component of total emissions, but again research in this
252 area is very limited. Effective modelling or up-scaling of GHG emissions from watershed to
253 regional/national scales is important for the improvement of global GHG budgets. Such up-
254 scaling needs an accurate estimation of C and N inputs and outputs i.e., a balance coupled with
255 net GHG emissions, while considering all possible processes and pathways involved. A study of
256 the dynamics of C and N in CWs is crucial, as the forms of removed C and N are particularly
257 pertinent to their potential for pollution swapping, global warming and water pollution.

258

259 Processes involved in N removal and N transformations in wetlands include sedimentation of
260 particulates (Koskiaho, 2003); nitrification, denitrification and DNRA (Poach et al., 2003; Burgin
261 et al., 2013), microbial assimilation and plant uptake-release (Findlay et al., 2003), anammox and
262 deamox (DENitrifying AMmonium OXidation). Constructed wetlands are complex systems that
263 facilitate aerobic and anaerobic microsites. Nitrification, denitrification and nitrifier
264 denitrification are the processes responsible for the production of N₂O. Depending on the
265 environmental conditions or management practices prevailing, a certain process will dominate
266 e.g. denitrification is the dominant process in SF CWs (Beaulieu et al., 2011), but nitrifier
267 denitrification is dominant in VSSF CWs (Wunderlin et al., 2013). Generally, CWs are anaerobic
268 but aquatic macrophytes can transport oxygen from the atmosphere to the rooting zone, where it

269 can sustain nitrification. The existence of microsites that support high activity and promote
270 denitrification has been shown in soils (Parkin, 1987) and such conditions are also likely to occur
271 in CWs where patchy distribution of organic material (e.g. particulate organic carbon) can occur
272 (Hamersley and Howes, 2002), which may be supplied by rhizodepositions (Minett et al., 2013).
273 Minett et al. (2013) found that simultaneous oxygenation of the rhizosphere, through radial
274 oxygen loss, and enhanced oxygen consumption by the soil occurs in the area immediately
275 surrounding the roots. Nitrate produced in the rooting zone can be taken up by plants or
276 denitrified and/or converted back to NH_4^+ by DNRA.

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278 Competition for NO_3^- may occur between denitrification and biotic assimilation. This is likely
279 governed by the prevailing aerobic/anaerobic conditions and therefore dependent on the type of
280 wetland. For instance in stormwater biofiltration systems, prolonged periods of inundation and
281 dry periods may support bio-assimilation over denitrification (Payne et al., 2014a,b).

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283 The conditions that favour the occurrence of either denitrification or DNRA are still in debate
284 (Rütting et al., 2011). DNRA is thought to be favoured by a C: NO_3^- ratio of >12 (Rütting et al.,
285 2011) and occurs at low levels of oxidation-reduction potential (Thayalakumaran et al., 2008).
286 The differences between denitrification and DNRA may be due to the availability of organic
287 matter, because DNRA is favoured at a high C: NO_3^- ratio and denitrification is favoured when
288 carbon supplies are limiting (Korom, 2002; Kelso et al., 1997). The fermentative bacteria that
289 carry out DNRA are obligate anaerobes, and so cannot occupy all the niches that denitrifiers can
290 (Buss et al., 2005). Takaya (2002) stated that a more reducing state favours DNRA over
291 denitrification. Pett-Ridge et al. (2006) showed that DNRA is less sensitive to dissolved oxygen
292 (DO) than denitrification. Fazzolari et al. (1998) showed that the effect of DO levels on DNRA is
293 dependent on the C: NO_3^- ratio and C, rather than DO, is the main factor regulating NO_3^-
294 partitioning between DNRA and denitrification. Significant DNRA may occur only at a C: NO_3^-
295 ratio above 12 (Yin et al., 1998). Different numbers of electrons are required in the reduction of
296 each NO_3^- molecule: five for denitrification and eight for DNRA. Therefore, more organic matter
297 can be oxidized for each molecule of NO_3^- by DNRA than by denitrification. In addition, NO_3^-
298 reduction is generally performed by fermentative bacteria that are not dependent on the presence
299 of NO_3^- for growth under anaerobic conditions. So, DNRA bacteria may be favoured by NO_3^- -
300 limited conditions (Laanbroek, 1990). Recent studies have suggested that DNRA may be an
301 important process compared to denitrification in wetland sediments (Burgin and Hamilton, 2008).
302 Van Oostrom and Russell (1994) found a 5% contribution of DNRA to the NO_3^- removal in CWs.

303 Little is known about the eventual fate of the NO_3^- that is converted to NH_4^+ via DNRA
304 pathways. In recent years, N cycling studies have increasingly investigated DNRA in various
305 ecosystems to explore its importance in N cycling (Rütting et al., 2011), but controls on DNRA
306 are relatively unknown (Burgin et al., 2013), DNRA being probably the least studied process of N
307 transformation in wetlands (Vymazal, 2007). However, DNRA can be a significant pathway of
308 NO_3^- reduction that impacts on the CW ecosystem services and so should therefore be evaluated.

309
310 Denitrification has been estimated to be a significant N removal process, but actual quantification
311 data are scarce. Few studies have estimated N losses by denitrification e.g. 19% (Mander et al.,
312 2008) and 86% (Obarska-Pempkowiak and Gajewska, 2003) of the total N input based on the
313 mass balance study. To our knowledge, no data are available on denitrification measurements in
314 soil/subsoils of surface flow CWs. While many of these pathways transfer Nr (mainly NH_4^+ and
315 N_2O) to the environment, other pathways can convert Nr to N_2 (e.g. denitrification, anammox and
316 deamox). Anammox can remove NO_2^- and NH_4^+ as N_2 when the existing environment is hypoxic.
317 Deamox can remove NO_3^- and NH_4^+ as N_2 , where NO_3^- is converted to NO_2^- by autotrophic
318 denitrification with sulphide (Kalyuzhnyi et al., 2006). In CWs, anammox and Deamox are not
319 well understood, so it is crucial to identify which of the processes are occurring in a specific type
320 of CW and the rate at which they occur. Once a process that provides N_2 as the end product is
321 determined, then the management of the CW could be directed towards enhancement of that
322 process. Hence, quantifying the rates of these processes for various types of CW is required for
323 improved N management towards lowering Nr in the environment.

324
325 The various components of the C cycle include: fixation of C by photosynthesis, respiration,
326 fermentation, methanogenesis and CH_4 oxidation with reduction of sulphur, iron and NO_3^- .
327 Anaerobic methane oxidation coupled with denitrification, a recently proposed pathway of the C
328 cycle (á Norði and Thamdrup, 2014; Haroon et al., 2013; Islas-Lima et al., 2004), can reduce CH_4
329 emissions in CWs. The C removal processes are sedimentation, microbial assimilation, gaseous
330 emissions, dissolved C losses through water to ground and surface water bodies, and chemical
331 fixation (bonding with chemical ions). Net primary productivity of wetland hydrophytes varies
332 across CW type, season, climatic region and local environmental conditions. For example, results
333 can vary remarkably for CWs containing the same plant species in different geographical regions
334 (Brix et al., 2001). Carbon mineralization in sediments depends on the redox chemistry of soil,
335 availability and quality of C (labile or recalcitrant) and temperature. In CWs, C cycling is very
336 complex due to the changes in redox chemistry, which regulates production and consumption of

337 CO₂ and CH₄ (Brix et al., 2001). In low redox conditions with limited DO, methanogens can
338 consume DOC and thus it is conducive to CH₄ production. The C:N ratios of wastewater affect
339 microbial growth and development which, in turn, affect their response to C and N cycles and
340 GHG emissions. Previous research on the effects of C:N ratios on nutrient removal and GHG
341 emissions are limited. A few examples include Yan et al. (2012) and Zhao et al. (2014), who
342 measured lower CO₂ and CH₄ emissions at C:N ratios of between 2.5:1 and 5:1, but this lower
343 range of C:N ratios decreased TOC removal. Hence, investigation of the influence of C:N ratio
344 on nutrient removal efficiencies and GHG emissions across CW and management types is crucial.
345 In summer, oxygen diffusion to the topsoil can reduce methanogenesis and stimulate CH₄
346 oxidation (Grünfeld and Brix, 1999). However, an increase in temperature can decrease DO in
347 deeper subsoil layers, which can enhance CH₄ production. Only a limited number of studies have
348 considered CH₄ and CO₂ efflux from CWs (e.g. Mander et al., 2008). As in all biochemical
349 reactions, temperature increases C and N turnover in CWs, causing high variations in GHG
350 emissions in different regions (temperate/ tropical/ arctic). These variations need to be considered
351 while extrapolating GHG emissions for different types of CWs under different management
352 practices.

353

354 Emissions of GHG in CWs can vary across CW typologies e.g. surface flow or subsurface flow
355 (Van der Zaag et al., 2010). A summary of N₂O, and CO₂ and CH₄ were presented in Table 3 and
356 Table 4, respectively. Generally, CH₄ emissions are higher in SF CWs than in SSF CWs (Table
357 3), but may vary with season. Nitrous oxide and CO₂ emissions are higher in VSSF CWs than
358 HSSF and SF CWs. The N₂O emissions factors (EF; N₂O/TN input ×100) ranged from 0.61 ±
359 0.21% in SF CWs to 1.01 ± 0.48% in VSSF CWs. The EF for CH₄ emissions ranged from 1.27 ±
360 0.31% in VSSF CWs to 16.8 ± 3.8% in SF CWs. The GHG from CWs can vary between
361 vegetated and non-vegetated systems (Table 5).

362

363 Aquatic plants play an important role in GHG production and transport to the atmosphere by
364 releasing GHG through their interconnected internal gas lacunas (Laanbroek, 2010). Emergent
365 plants can transport atmospheric oxygen to the rooting zone and contribute to increased N₂O and
366 CO₂ production and CH₄ consumption (Brix, 1997). Vascular plants can exchange GHG between
367 the rooting zone and atmosphere (Yavitt and Knapp, 1998). Vegetation and its composition affect
368 the nutrient dynamics and the production, consumption and transport of GHG and hence their
369 exchange between wetlands and atmosphere (Ström et al., 2003, 2005; Søvic et al., 2006;
370 Johansson et al., 2003). They can also affect the biogeochemistry of CWs due to the differences

371 in their growth and development, longevity, root systems, root density, root depth and microbial
372 ecology in the rhizosphere. As some plant litter decomposes, organic matter with lignocellulose
373 and humic compounds may be released that are more or less labile or stable in nature than others.
374 Release of low molecular weight organic matter that is labile in nature is more likely to produce
375 GHGs than stable forms. For example, *Z. latifolia* showed higher nutrient removal and CH₄
376 fluxes than *P. australis* (Inamori et al., 2007). The *Z. lotifolia* root system is shallow and the
377 activity of methanotrophs is primarily confined to the top soil. The root systems of *P. australis*
378 are deeper, which is more favourable for the oxidation of CH₄. A fluctuating water table in CWs
379 has significant impacts on GHG dynamics. Pulsing hydrologic regimes decreases CH₄ but
380 increases N₂O emissions. In aerobic and anaerobic conditions caused by pulsing hydrology,
381 incomplete nitrification and denitrification increase N₂O emissions. However, the effects of
382 pulsing hydrologic regimes on GHG emissions are contradictory. For example, intermittent
383 hydrologic regimes decrease both N₂O (Sha et al., 2011) and CH₄ emissions (Song et al., 2010).
384 Highly contrasting results on gas emissions with fluctuating water levels have been reported and
385 the controlling mechanisms are unclear (Elberling et al., 2011).

386

387 Therefore, the assessment of GHG emissions in various types of CW (surface flow, subsurface
388 flow, vertical and horizontal), vegetation cover (vegetated, nonvegetated) and species type, and
389 management system employed (HLR, HRT, soil used and water table), is necessary in light of the
390 national and global GHG budgets. In addition, such measurements will help scientists,
391 environmental managers and policy makers to adopt environmentally friendly construction and
392 management of CWs. The enhanced reduction of N₂O to N₂ needs further elucidation.

393

394 5. Surface Emissions vs Subsurface Export of C and N

395 Dissolved GHG produced in soils and subsoils can be emitted to atmosphere by transpiration of
396 vascular plants (from within the rooting zone), ebullition and diffusion from soils. Elberling et al.
397 (2011) reported that in wetlands, the transport of gases through subsoil occurs both *via* diffusive
398 transport in the pores and through the vascular plants. Surface emissions of GHG from CWs are
399 well recognised and have been commonly measured by chamber methods. The GHG produced in
400 CWs can also be transported to the groundwater with the percolating water and emitted to the
401 atmosphere upon discharge to surface waters. It can also flow towards surface waters by
402 advective transport and/or by dispersion of groundwater. Dissolved nutrients can be preferentially
403 leached down into deeper soil layers and groundwater *via* different pathways (e.g. root channels).
404 The Nr delivered to groundwater can be transformed *in situ* to other reactive or benign forms.

405 Hence, quantification of such Nr loadings to groundwater and their *in situ* consumption (e.g. N₂O
406 to N₂ or CH₄ to CO₂) is necessary to understand their environmental consequences. In addition,
407 DON, NO₃⁻ and NH₄⁺ and DOC delivered to surface waters can undergo biochemical reactions
408 and produce N₂O, CO₂ and CH₄ in streams and estuaries. Ström et al. (2007) measured a
409 considerable quantity of CH₄ in porewater and found a correlation between the surface emissions
410 and porewater CH₄ concentrations in vegetated wetlands. Measuring only the surface emissions
411 of GHG can omit substantial quantities of GHG released from CWs. For example, Riya et al.
412 (2010) measured emissions of CH₄ and N₂O accounting for 2.9 and 87% of the total emissions.
413 Measuring porewater GHG and linking these to the surface emissions and subsurface export to
414 groundwater below CWs will help to estimate a better GHG balance from both a national and
415 global context. Elberling et al. (2011) linked subsurface gas concentrations in wetlands to the
416 surface fluxes using a diffusion model. This demonstrates the need for future studies on
417 subsurface GHG production, consumption and net GHG emissions in CWs within a climate
418 change context.

419

420 It is important to characterise soils and subsoils physical (e.g. texture, bulk density) and hydraulic
421 (development of a soil water characteristic curve) properties and to assess their potential to
422 percolate dissolved nutrients and gases in the solute phase to the underlying groundwater. To our
423 knowledge, the indirect pathway of GHG emissions from CWs has never been reported, despite
424 the fact that this would appear to have a high biogeochemical potential to produce and exchange
425 GHG. The balance between N and C input and output flows between CWs and aquatic and
426 atmospheric environments, together with the direct and indirect emissions of C and N species,
427 could be an important input to global C and N budgets.

428

429 6. Hydrogeochemistry below CWs

430 Constructed wetlands can be designed with or without a clay liner or a compacted soil bed at the
431 base, which can lead to large differences in permeability of the underlying layers. The variation
432 in permeability of a CW soil bed will affect solute, nutrient and GHG flows, and their interactions
433 with the underlying groundwater (Dzakpasu et al., 2012; 2014). Groundwater hydrogeochemistry
434 below CWs can therefore provide a unique insight into such interactions. An example of such
435 interactions would be between nutrient-rich water discharging from CW cells mixing with
436 laterally moving regional groundwater. It should be noted that groundwater can also discharge
437 into CWs depending on the hydraulic gradients. This means that fully screened, multi-level

438 piezometers or boreholes should be installed at such sites to elucidate groundwater flow direction,
439 hydraulic gradients and conductivities. Such monitoring networks allow water samples to be
440 collected and the sources of nutrients in groundwater bodies below CWs to be identified. The
441 local site hydrology (precipitation, groundwater table fluctuations and evapotranspiration) has a
442 large impact on the pollutant removal. Hydrogeochemical studies at an accurate spatial and
443 temporal resolution should explain the effects of precipitation on nutrient removal by dilution as
444 well *in situ* nutrient turnover. Effective CW management requires an understanding of the effects
445 of wetland hydrology on the physical and biochemical attenuation of nutrients in order to assess
446 their impacts on the surface emissions and subsurface export of nutrients and GHG. Data on the
447 species of N in groundwater below the CWs are required to provide an in-depth understanding of
448 wetland ecosystem services, particularly if CWs have the potential to leak pollutants down into
449 the groundwater (Dzakpasu et al., 2014). Higher NH_4^+ concentrations in groundwater below the
450 CWs than the effluent are often reported (Harrington et al., 2007; Dzakpasu et al., 2012).
451 Therefore, questions arise with respect to NH_4^+ concentrations in groundwater below the CWs if
452 they have been transported from CWs. Linking geochemistry of groundwater below CWs to site
453 hydrology, water table fluctuations and soil/subsoil physico-chemical properties is required to
454 elucidate the major environmental drivers of C and N removal, and/or pollution swapping. The
455 quality of groundwater underlying CWs with regards to the Nr species is largely unknown.

456

457 7. Methodological Developments

458 To improve the ecosystem services and to minimize the pollution swapping of CWs,
459 quantification of N cycling is crucial. Measurement of GHG using the closed chamber method is
460 widely used, but has large uncertainty in estimating the diurnal variability due to internal changes
461 in temperature and physical access to the chambers over a 24-h time period. Gas ebullition and
462 diffusion measurements are quite challenging in CWs covered by vegetation, because of the
463 difficulties in estimation of gas transfer velocity. Application of the eddy covariance method is
464 not appropriate for most CWs, as it requires a large surface area (> several ha) to avoid
465 contribution of surrounding area and complication of GHG foot printing. A combination of
466 chamber, ebullition and diffusion methods in a single system could minimise the uncertainty in
467 GHG estimation. Methane ebullition measurement was found to be similar to surface emissions
468 by the chamber method, but N_2O and CO_2 ebullition measurements were lower than the surface
469 emissions (Søvik et al., 2006).

470

471 The use of *in situ* microcosm study and soil core incubation methods may give a better estimation
472 of N₂O, CO₂ and CH₄ production and consumption than existing methods. With the recent
473 advancement of isotope pairing and dilution techniques, single or simultaneously occurring C and
474 N transformation processes can be quantified in laboratory or *in situ* conditions (Huygens et al.,
475 2013; Müller et al., 2014). The isotope technique relies on the introduction of a known amount of
476 ¹⁴C and or ¹⁵N into the CW and then quantification of C and N concentrations and isotopic
477 compositions through different C and N pools after incubation for a specific period. Laboratory
478 methods involve collection of intact soil/sediment cores, with subsequent incubation in the
479 laboratory. *In situ* field techniques involve release of ¹⁴C/ ¹⁵N solution in the CW soils.
480 Incubation of intact soil cores with differentially labelled ¹⁵NH₄¹⁴NO₃ and ¹⁴NH₄¹⁵NO₃ can be
481 used to quantify the rates of different N transformation processes (Rütting and Müller, 2008). The
482 quantification of simultaneously occurring N transformation rates rely on the analysis with
483 appropriate ¹⁵N tracing models. In recent years, ¹⁵N tracing techniques have evolved, and are now
484 able to identify process-specific NO₂⁻ pools (Rütting and Müller 2008), pathway-specific N₂O
485 production and emission, as well as N₂O:N₂ ratios (Müller, Laughlin et al. 2014). Traditional
486 techniques for investigation of gross N dynamics in sediments (Blackburn 1979) may be
487 combined with the latest ¹⁵N tracing techniques, where all N transformation rates are included
488 (Huygens, Trimmer et al. 2013). Thus, current models should consider processes such as
489 anammox and/or deammox, and then be tested in CWs under various operational conditions.
490 Denitrification in porewater samples can be measured by analysing samples for dissolved N₂ in a
491 membrane inlet mass spectrometer (MIMS; Kana et al., 1994) and N₂O in a gas chromatograph
492 (GC; Jahangir et al., 2012). The studies of natural abundance of ¹⁵N and ¹⁸O (δ¹⁵N and δ¹⁸O) in
493 NO₃⁻ is an insightful tool for the investigation of the sources, fate and transformational processes
494 of N in a system (e.g. in shallow groundwater; Baily et al., 2011). The *in situ* NO₃⁻ push-pull
495 method has been used to determine denitrification in shallow groundwater (<3 m) in riparian
496 wetlands (Addy et al., 2002; Kellogg et al., 2005) and in deep groundwater in arable/grassland
497 (Jahangir et al., 2013).

498
499 Isotope-based techniques can also be extended to other elements e.g., a ³³P tracing model has
500 been developed recently to study phosphorus (P) cycle in soil (Müller and Bünemann, 2014).
501 These techniques can be applied in the study of C, N and P biogeochemistry in aquatic
502 environments. In addition, measurements of DOC and gases (CO₂ and CH₄) will provide insights
503 into the C consumption and transformation associated with the N transformations. Carbon and N
504 dynamics are influenced by the interacting effects of soil conditions with microbial community

505 structure and functioning. Microbial functioning involves transcription of genes, translation of
506 messenger RNA and activity of enzymes (Firestone et al., 2012). As such, activities of microbial
507 communities under various environmental conditions and how these contribute to C and N
508 dynamics is a very important area of future research (Müller and Clough, 2014). Molecular
509 approaches can be important tools for identifying and quantifying the genes that code for
510 enzyme-mediating C and N cycles (Peterson et al., 2012). These tools help assess the
511 relationships among genes, environmental controllers, and the rates of C and N processes. The
512 scientific tools and multidisciplinary techniques are now available to better understand C and N
513 transformation rates, processes and factors controlling the unwanted emission of N and C
514 products to the environment.

515

516 8. Conclusions and Recommendations

517 The transformational processes on a mixture of contaminants within and below CWs can cause
518 pollution swapping. A holistic assessment of C and N dynamics in CWs is needed to fully
519 understand their removal, transport and impact on water quality and emissions to atmosphere.
520 Mixed contaminants entering CWs and those formed within and underneath CWs during
521 transformational processes must be considered in future studies. The overall balance of these
522 constituents will decide on whether a CW is a pollution source or a sink. This will necessitate a
523 higher degree of multi-level spatial and temporal monitoring and the use of multi-disciplinary
524 techniques both *in* and *ex situ* to fully characterise all pathways of C and N loss. At this time we
525 cannot suggest any design optima in terms of nutrient removal and GHG mitigation because
526 empirical information are not yet abundant. To do this, transformation kinetics of C and N and
527 net GHG emissions through all possible pathways are required to provide a holistic assessment.
528 However, a combination of various types of CW and plant types could provide higher removals
529 and lower GHG emissions. A conceptual model highlighting the current state of knowledge in
530 this area and the research gaps is presented in Figure 1.

531

532 Subsurface export of nutrients and GHG to groundwater should be accounted for in CW
533 management. Reducing the saturated hydraulic conductivity below the wetland bed will help
534 reduce nutrients leaching to groundwater. The reactive versus the benign forms of the N
535 transformation products should be evaluated. Data on when, where and the rates at which
536 denitrification, deamox and anammox occur in CWs are needed, as well as identification of the
537 key factors that control such processes. The provenance of NH_4^+ in groundwater below CW cells

538 and its impact on down-gradient receptors needs further elucidation. Constructed wetlands have
539 the potential to produce N₂O, DON, DOC, dissolved inorganic C (DIC), CO₂ and CH₄, which
540 may be exported to fresh waters *via* groundwater and degassed upon discharge to surface waters.
541 Moreover, the DOC and DIC transferred to the fresh water sediments (rivers and lakes) can
542 produce GHG that, in turn, emit to atmosphere. The amount of C and N exported from terrestrial
543 ecosystems *via* the subsurface pathway to fresh waters has been the missing piece of our
544 understanding of global C and N budgets. It is clear that data on the various C and N species,
545 along with the GHG emissions, are crucial to make a robust input-output balance of C and N in
546 CWs. Spatial and temporal variations of GHG emissions in CWs under different management
547 systems are also critical to get much more rigorous estimates of emission factors. These data will
548 reduce the existing uncertainties in global C and N budgets.

549

550 Managing wetting and drying spells (pulsing hydrology) in CWs can enhance NH₄⁺ removal.
551 Similarly, oxidation of organic C will increase CO₂ production and, in anaerobic conditions, may
552 be reduced to CH₄. This requires more research into the C and N cycle processes over the wetting
553 and drying spells, which is now possible with the advancement in ¹⁴C/¹⁵N tracing and modelling
554 techniques. The selection of appropriate plant species is important to optimise nutrient removal,
555 sequester C and decrease GHG emissions, but more research is needed across species and
556 geographical locations. Further research is also needed to investigate the impacts of hydraulic
557 retention time on nutrient dynamics. Rates of nutrient accumulation or fixation in soils and their
558 *in situ* transformation in CWs need to be quantified to evaluate their contribution to C
559 sequestration and GHG emissions.

560

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564

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574 References

575 á Norði, K. and Thamdrup, B.: Nitrate-dependent anaerobic methane oxidation in a freshwater
576 sediment, *Geochimica et Cosmochimica Acta*, 132, 141-150, 2014.

577 Addy, K., Kellogg, D.Q., Gold, A.J., Groffman, P.M., Ferendo, G., and Sawyer, C.: *In situ* push-
578 pull method to determine groundwater denitrification in riparian zones, *J. Environ. Qual.*,
579 31, 1017-1024, 2002.

580 Bachand, P.A.M., and Horne, A.J.: Denitrification in constructed free-water surface wetlands: II.
581 Effects of vegetation and temperature, *Ecol. Eng.*, 14, 17–32, 2000.

582 Baily, A., Rock, L., Watson, C.J., and Fenton, O.: Spatial and temporal variations in groundwater
583 nitrate at an intensive dairy farm in south-east Ireland: Insights from stable isotope data,
584 *Agril. Ecosyst. Environ.*, 308-318, 2011.

585 Beaulieu, J.J., Tank, J.L., Hamilton, S.K., Wollheim, W.M., Hall, R.O., and Mulholland, P.J.:
586 Nitrous oxide emission from denitrification in stream and river networks, *Proc. Natl.*
587 *Acad. Sci.*, 108, 214–219, 2011.

588 Blackburn, T.H.: Methods for measuring rates of NH_4^+ turnover in anoxic marine sediments,
589 using a $^{15}\text{N-NH}_4^+$ dilution technique, *Appl. Environ. Microbiol.*, 37(4), 760-765, 1979.

590 Bojcevska, H., and Tonderski, K.: Impact of loads, season, and plant species on the performance
591 of a tropical constructed wetland polishing effluent from sugar factory stabilization ponds,
592 *Ecol. Eng.*, 29, 66–76, 2007.

593 Brix, H.: Do macrophytes play a role in constructed treatment wetlands, *Water Sci. Technol.*, 35,
594 11-17, 1997.

595 Burgin, A.J., Hamilton, S.K., Gardner, W.S., and McCarthy, M.J.: Nitrate reduction,
596 denitrification, and dissimilatory nitrate reduction to ammonium in wetland sediments, in:
597 R.D. DeLaune, K.R. Reddy, C.J. Richardson, and J.P. Megonigal, (Eds.), *Methods in*
598 *Biogeochemistry of Wetlands*, SSSA Book Series, no. 10, Madison, USA, 307-325, 2013.

599 Calheiros, C.S.C., Rangel, A.O.S.S., and Castro, P.M.L.: Constructed wetland systems vegetated
600 with different plants applied to the treatment of tannery wastewater, *Water Res.* 41, 1790–
601 98, 2007.

602 Clair, T.A., Arp, P., Moore, T.R., Dalva, M., and Meng, F.R.: Gaseous carbon dioxide and
603 methane, as well as dissolved organic carbon losses from a small temperate wetland under
604 a changing climate, *Environ. Pollut.*, 116(1), S143-S148, 2002.

605 Conley, D.J., Paerl, H.W., Howarth, R.W. Boesch, D.F., Seitzinger, S.P., Havens, K.E., Lancelot,
606 C., and Likens, G.E.: Controlling eutrophication: Nitrogen and phosphorus, *Science*, 323,
607 1014–1015, 2009.

608 da Motta Marques D.M.L., Leite, G.R., and Giovannini, S.G.T.: Performance of two macrophyte
609 species in experimental wetlands receiving variable loads of anaerobically treated
610 municipal wastewater, *Water Sci. Technol.*, 44, 311–6, 2001.

611 Dunne, E.J., Culleton, N., O'Donovan, G., Harrington, R., and Olsen, A.E.: An integrated
612 constructed wetland to treat contaminants and nutrients from dairy farmyard dirty water,
613 *Ecol. Eng.*, 24(3), 219-232, 2005.

614 Dzakpasu, M., Scholz, M., Harrington, R., McCarthy, V. and Jordan, S.: Groundwater Quality
615 Impacts from a Full-Scale Integrated Constructed Wetland, *Groundwater Monit. Rem.*,
616 34(3), 51-64, 2014.

617 Dzakpasua, M., Scholz, M., Harrington, R., Jordan. S.N., and McCarthy, V.: Characterising
618 infiltration and contaminant migration beneath earthen-lined integrated constructed
619 wetlands, *Ecol. Eng.*, 41, 41-51, 2012.

620 Elberling, B., Louise A., Christian, J.J., Hans, P.J., Michael K., Ronnie N.G., and Frants, R.L.:
621 Linking soil O₂, CO₂, and CH₄ concentrations in a wetland soil: implications for CO₂ and
622 CH₄ fluxes, *Environ. Sci. Technol.*, 45, 3393–3399, 2011.

623 Fazzolari, C. E., Nicolardot, B., and Germon, J. C.: Simultaneous effects of increasing levels of
624 glucose and oxygen partial pressures on denitrification and dissimilatory reduction to
625 ammonium in repacked soil cores, *Eur. J. Soil Biol.*, 34, 47–52, 1998.

626 Fenton, O., Healy, M.G., Brennan, F., Jahangir, M.M.R., Lanigan, G.J., Richards, K.G.,
627 Thornton, S.F., and Ibrahim, T.G.: Permeable reactive interceptors – blocking diffuse
628 nutrient and greenhouse gas losses in key areas of the farming landscape, *J. Agric. Sci.*,
629 152, S71-S81, 2014.

630 Fey, A., Benckiser, G., and Ottow, J.C.G.: Emissions of nitrous oxide from a constructed wetland
631 using a ground filter and macrophytes in waste-water purification of a dairy farm, *Biol.*
632 *Fertil. Soils*, 29(4), 354–359, 1999.

633 Findlay, S., Groffman, P., and Dye, S.: Effects of *Phragmites australis* removal on marsh nutrient
634 cycling, *Wetlands Ecol. Manage.*, 11, 157-165, 2003.

635 Firestone, M., Blazewicz, S., Peterson, D.G., and Placella, S.: Can molecular microbial ecology
636 provide new understanding of soil nitrogen dynamics, in: Richards, K.G., Fenton, O.,
637 Watson, C. J. (Eds.), *Proceedings of the 17th Nitrogen Workshop*, 11-14, 2012.

638 Galloway, J.N., Aber, J.D., Erisman, J.W., Seitzinger, S.P., Howarth, R.W., Cowling, E.B., and
639 Cosby, B.J.: The nitrogen cascade, *BioSci.*, 53(4), 341-356, 2003.

640 Garcia, J., Capel, V., Castro, A., Ruiz, I., and Soto, M.: Anaerobic biodegradation tests and gas
641 emissions from subsurface flow constructed wetlands, *Bioresour. Technol.*, 98(16), 3044–
642 3052, 2007.

643 Gold, A.J., Eddy, K., David, M.B., Schpper L.A., and Needelman, B.A.: Artificial sinks:
644 opportunities and challenges for managing offsite nitrogen losses, *J. Contemp. Water Res.*
645 *Edu.*, 151, 9-19, 2013.

646 Gray, N.F.: *Drinking water quality*, 2nd edn., Cambridge Univ. Press, Cambridge, UK, 2008.

647 Grünfeld, S., and Brix, H.: Methanogenesis and methane emissions: effects of water table,
648 substrate type and presence of *Phragmites australis*, *Aquatic Bot.*, 64, 63-75, 1999.

649 Gui, P., Inamori, R., Matsumura, M., and Inamori, Y.: Evaluation of constructed wetlands by
650 waste water purification ability and greenhouse gas emissions, *Water Sci. Technol.*, 56(3),
651 49–55, 2007.

652 Haroon, M. F., Hu, S., Shi, Y., Imelfort, M., Keller, J., Hugenholtz, P., Yuan, Z., and Tyson,
653 G.W.: Anaerobic oxidation of methane coupled to nitrate reduction in a novel archaeal
654 lineage, *Nature*, 500, 567-570, 2013.

655 Harrington, R., Carroll, P., Carty, A.H., Keohane, J., and Ryder, C.: Integrated constructed
656 wetlands: concept, design, site evaluation and performance, *Int. J. Water*, 3(3), 243-255,
657 2007.

658 Healy, M.G., Ibrahim, T.G., Lanigan, G., Serrenho, A.J., and Fenton, O.: Nitrate removal rate,
659 efficiency and pollution swapping potential of different organic carbon media in
660 laboratory denitrification bioreactors, *Ecol. Eng.*, 40, 198-209, 2011.

661 Healy, M.G., Barrett, M., Lanigan, G.J., João Serrenho, A., Ibrahim, T.G., Thornton, S.F., Rolfe,
662 S.A., Huang, W.E., Fenton, O.: Optimizing nitrate removal and evaluating pollution
663 swapping trade-offs from laboratory denitrification bioreactors, *Ecol. Eng.*, 74, 290-301,
664 2014

665 Hill, A.R.: Nitrate removal in stream riparian zones, *J. Environ. Qual.*, 25, 743-755, 2010.

666 Huygens, D., Trimmer, M., Rütting, T., Müller, C., Heppell, C.M., Lansdown, K., and Boeckx,
667 P.: Biogeochemical N cycling in wetland ecosystems: ¹⁵N isotope techniques, *Methods in*
668 *biogeochemistry of wetlands*, K.R. Reddy, J.P. Megonigal and R.D. Delaune, (Eds.), *Soil*
669 *Sci. Soc. Amer.*, 30, 553-591, 2013.

670 Inamori, R., Gui, P., Dass, P., Matsumura, M., Xu, K.Q., Kondo, K., Ebie, Y., and Inamori, Y.:
671 Investigating CH₄ and N₂O emissions from eco-engineering wastewater treatment
672 processes using constructed wetland microcosms, *Proc. Biochem*, 42, 363-373, 2007.

673 IPCC: 2013 Supplement to the 2006 IPCC guidelines for national greenhouse gas inventories:
674 wetlands, edited by: Hirashi, T., Krug, T., Tanabe, K., Srivastava, N., Baasansuren, J.,
675 Fukuda, M., Troxler, T.G., Switzerland, IPCC Task Force on National Greenhouse Gas
676 Inventories, 354, 2014.

677 Islas-Lima, S., Thalasso, F., and Gomez-Hernandez, J.: Evidence of anoxic methane oxidation
678 coupled to denitrification, *Water Res.*, 38(1), 13-16, 2004.

679 Jahangir, M.M.R., Johnston, P., Addy, K., Khalil, M.I., Groffman, P., and Richards, K.G.:
680 Quantification of in situ denitrification rates in groundwater below an arable and a
681 grassland system, *Water Air Soil Pollut.*, 224, 1693, doi:10.1007/s11270-013-1693-z,
682 2013.

683 Jahangir, M.M.R., Johnston, P., Grant, J., Somers, C., Khalil, M.I., and Richards, K.G.:
684 Evaluation of headspace equilibration methods for measuring greenhouse gases in
685 groundwater, *J. Environ. Manage.*, 111, 208-212, 2012.

686 Johansson, A.E., Gustavsson, A.M., Öquist, M.G., and Svensson, B.H.: Methane emissions from
687 a constructed wetland treating wastewater – seasonal and spatial distribution and
688 dependence on edaphic factors, *Water Res.*, 38 (18), 3960–3970, 2004.

689 Johansson, A.E., Kasimir-Klemedtsson, A., Klemedtsson, L., and Svensson, B.H.: Nitrous oxide
690 exchanges with the atmosphere of a constructed wetland treating wastewater, *Tellus*, 55B,
691 737–750, 2003.

692 Johansson, E.: Constructed wetlands and deconstructed discourses—greenhouse gas fluxes and
693 discourses on purifying capacities, *Dept. Water Environ. Studies, Linköping Univ.*,
694 Sweden, 2002.

695 Kadlec, R. H., and Wallace, S. D.: *Treatment wetlands*, CRC Press Taylor and Francis, 1000 pp.,
696 2009.

697 Kalyuzhnyi, S., Gladchenko, M., Mulder, A., and Versprille, B.: DEAMOX-new biological
698 nitrogen, *Water Res.*, 40, 3637-3645, 2006.

699 Kana, T.M., Darkangelo, C., Hunt, M.D., Oldham, J.B., Bennett, G.E, and Cornwell, J.C.:
700 Membrane inlet mass spectrometer for rapid high precision determination N₂, O₂ and Ar
701 in environmental water samples, *Anal. Chem.*, 66, 4166-4170, 1994.

702 Kato, K., Koba, T., Ietsugu, H., Saigusa, T., Nozoe, T., Kobayashi, S., Kitagawa, K., and
703 Yanagiya, S.: Early performance of hybrid reed bed system to treat milking parlour

704 wastewater in cold climate in Japan, in: 10th international conference wetland systems for
705 water pollution control, MAOTDR, Lisbon, Portugal, pp. 1111-1118, 2006.

706 Kellogg, D.Q., Gold, A.J., Groffman, P.M., Addy, K., Stolt, M.H., and Blazejewski, G.: *In situ*
707 groundwater denitrification in stratified, permeable soils underlying riparian wetlands, J.
708 Environ. Qual., 34, 524-533, 2005.

709 Kelso, B.H.L., Smith, R.V., Laughlin, R.J., and Lennox, S.D.: Dissimilatory nitrate reduction in
710 anaerobic sediments leading to river nitrate accumulation, Appl. Environ. Microbiol.,
711 63(12), 4679-4685, 1997.

712 Korom, S.F.: Natural denitrification in the saturated zone: a review, Water Resour. Res., 28(6),
713 1657–1668, 2002.

714 Koskiaho, J., Ekholm, P., Rätty, M., Riihimäki, J., and Puustinen, M.: Retaining agricultural
715 nutrients in constructed wetlands- experiences under boreal conditions, Ecol. Eng., 20(1),
716 89–103, 2003.

717 Laanbroek, H.J.: Bacterial cycling of minerals that affect plant growth in waterlogged soils: a
718 review, Aquat. Bot., 38, 109–25, 1990.

719 Laanbroek, H.J.: Methane emission from natural wetlands: interplay between emergent
720 macrophytes and soil microbial processes, A mini-review, Annals Bot., 105, 141–153,
721 2010.

722 Langergraber, G.: Modeling of processes in subsurface flow constructed wetlands: A review.
723 Vadose Zone, J. 7, 830-842, 2008.

724 Lee, C., Fletcher, T.D., and Sun, G.: Nitrogen removal in constructed wetland systems, Eng. Life
725 Sci., 9(1), 11-22, 2009.

726 Liikanen, A., Huttunen, J.T., Kaijalainen, S.M., Heikkinen, K., Vaisänen, T.S., Nykiinen, H., and
727 Martikainen, P.J.: Temporal and seasonal changes in greenhouse gas emissions from a
728 constructed wetland purifying peat mining runoff water, Ecol. Eng., 26, 241-251, 2006.

729 Liu, C., Xu, K., Inamori, R., Ebie, Y., Liao, J., and Inamori, Y.: Pilot-scale studies of domestic
730 wastewater treatment by typical constructed wetlands and their greenhouse gas emissions,
731 Front. Environ. Sci. Eng. China, 3(4), 477–482, 2009.

732 Luederitz, V., Eckert, E., Lange-Weber, M., Lange, A., and Gersberg, R.M.: Nutrient removal
733 efficiency and resource economics of vertical flow and horizontal flow constructed
734 wetlands, Ecol. Eng., 18, 157–171, 2001.

735 Luo, W.G., Wang, S.H., Huang, J., and Qian, W.Y.: Denitrification by using subsurface
736 constructed wetland in low temperature. China Water Wastewater (in Chinese), 21, 37–
737 40, 2005.

738 Makino, W., Cotner, J.B., Sterner, R.W., and Elser, J.J.: Are bacteria more like plants or animals?
739 Growth rate and resource dependence of bacterial C:N:P stoichiometry, *Funct. Ecol.*, 17,
740 121–130, 2003.

741 Maltais-Landry, G., Maranger, R., Brisson, J., and Chazarenc, F.: Greenhouse gas production and
742 efficiency of planted and artificially aerated constructed wetlands, *Environ. Pollut.*, 157,
743 748-754, 2009.

744 Mander, U., Löhmus, K., Teiter, S., Mairing, T., Nurk, K., and Augustin, J.: Gaseous fluxes in
745 the nitrogen and carbon budgets of subsurface flow constructed wetlands, *Sci. Total*
746 *Environ.*, 404, 343-353, 2008.

747 Mander, Ü., Löhmus, K., Teiter, S., Nurk, K., Mairing, T., and Augustin, J.: Gaseous fluxes from
748 subsurface flow constructed wetlands for wastewater treatment, *J. Environ. Sci. Health A*
749 *40(6–7)*, 1215–26, 2005.

750 Mander, Ü., Maddison, M., Soosaar, K., and Karabelnik, K.: The impact of intermit-tent
751 hydrology and fluctuating water table on greenhouse gas emissions from subsurface flow
752 constructed wetlands for wastewater treatment, *Wetlands*, 31(6), 1023–1032, 2011.

753 Minett, D.A., Cook, P.L.M., Kessler, A.J., and Cavagnaro, T.R.: Root effects on the spatial and
754 temporal dynamics of oxygen in sand-based laboratory-scale constructed biofilters, *Ecol.*
755 *Eng.*, 58, 414-422, 2013.

756 Mitsch, W.J., and Gosselink, J.G.: *Wetlands*, 3rd Edn. New York, John Wiley & Sons, p. 936,
757 2000.

758 Müller, C. and Bünemann, E.K.: A ³³P tracing model for quantifying gross P transformation rates
759 in soil, *Soil Biol. Biochem.*, 76, 218-226, 2014.

760 Müller, C. and Clough, T.J.: Advances in understanding nitrogen flows and transformations: gaps
761 and research pathways, *J. Agril. Sci.*, 152, S1, 34-44, 2014.

762 Müller, C., Laughlin, R.J., Spott, O., and Rütting, T.: Quantification of N₂O emission pathways
763 via a ¹⁵N tracing model, *Soil Biol. Biochem.*, 72, 44-54, 2014.

764 Mustafa, A., and Scholz, M.: Nutrient accumulation in *Typha latifolia* L. and sediment of a
765 representative integrated constructed wetland, *Water Air Soil Pollut.*, 219, 329-341, 2011.

766 Nguyen, L.M.: Organic matter composition, microbial biomass and microbial activity in gravel-
767 bed constructed wetlands treating farm dairy wastewaters, *Ecol. Eng.*, 16, 199-221, 2000.

768 O’Luanaigh, N.D., Goodhue, R., and Gill, L.W.: Nutrient removal from on-site domestic
769 wastewater in horizontal subsurface flow reed beds in Ireland, *Ecol. Eng.*, 36, 1266-1276,
770 2010.

771 Obarska-Pempkowiak, H., and Gajewska, M.: The dynamics of processes responsible for
772 transformation of nitrogen compounds in hybrid wetlands systems in a temperate climate,
773 in: J. Vymazal (Eds.) *Wetlands- Nutrients, Metals and Mass Cycling*. Backhuys
774 Publishers, Leiden, The Netherlands, p.129–42. 2003.

775 Pangala, S.R., Reay, D.S., and Heal, K.V.: Mitigation of methane emissions from constructed
776 farm wetlands, *Chemosphere*, 78, 493-499, 2010.

777 Payne, E. G. I., Fletcher, T. D., Cook, P. L. M., Deletic, A., and Hatt, B. E.: Processes and drivers
778 of nitrogen removal in stormwater biofiltration, *Critical Reviews in Environmental*
779 *Science and Technology*, 44, 796-846, 2014a.

780 Payne, E. G. I., Fletcher, T. D., Russell, D. G., Grace, M. R., Cavagnaro, T. R., Evrard, V.,
781 Deletic, A., Hatt, B. E., and Cook, P. L. M.: Temporary storage or permanent removal?
782 The division of nitrogen between biotic assimilation and denitrification in stormwater
783 biofiltration systems, *PLoS ONE*, 9, e90890, 2014b.

784 Peterson, D.G., Blazewicz, S., Herman, D.J., Firestone, M., Turetsky, M. and Waldrop, M.:
785 Abundance of microbial genes associated with nitrogen cycling as indices of
786 biogeochemical process rates across a vegetation gradient in Alaska, *Environ. Microbiol.*,
787 14, 993-1008, 2012.

788 Pett-Ridge, J., Silver, W. L., and Firestone, M.K.: Redox fluctuations frame microbial community
789 impacts on N-cycling rates in humid tropical forest soil, *Biogeochem.*, 81, 95–110, 2006.

790 Poach, M.E., Hunt, P.G., Vanotti, M.B., Stone, K.C., Matheny, T.A., Johnson, M.H., and Sadler
791 E.J.: Improved nitrogen treatment by constructed wetlands receiving partially nitrified
792 liquid swine manure, *Ecol. Eng.*, 20, 183-197, 2003.

793 Rabalais, N.N., Diaz, R.J. Levin, L.A. Turner, R.E., Gilbert, D., and Zhang, J.: Dynamics and
794 distribution of natural and human-caused coastal hypoxia, *Biogeosci.*, 7, 585–619, 2010.

795 Reay, D.S.: Title: Fertilizer 'solution' could turn local problem global - Protecting soil and water
796 from pollution may mean releasing more greenhouse gas, *Nature*, 427, 485-485, 2004.

797 Reddy, K. R., and Delaune, R. D.: *Biogeochemistry of Wetlands: Science and Applications*, CRC
798 Press, 800 pp., 2008.

799 Riya, S., Zhou, S., Nakashima, Y., Terada, A., Hosomi, M.: Direct and indirect greenhouse gas
800 emissions from vertical flow constructed wetland planted with forage rice, *Kagaku*
801 *Kogaku Ronbunshu*, 36, 229–236, 2010.

802 Rütting, T. and Müller, C.: Process-specific analysis of nitrite dynamics in a permanent grassland
803 soil by using a Monte Carlo sampling technique, *Euro. J. Soil Sci.*, 59: 208-215, 2008.

804 Rütting, T., Boeckx, P., Müller, C., and Klemetsson, L.: Assessment of the importance of
805 dissimilatory nitrate reduction to ammonium for the terrestrial nitrogen cycle, *Biogeosci.*,
806 8, 1169-1196, 2011.

807 Seitzinger, S.P., Sanders, R.W., and Styles, R.: Bioavailability of DON from natural and
808 anthropogenic sources to estuarine plankton, *Limnol. Oceanogr.*, 47, 353–366, 2002.

809 Sha, C.Y., Mitsch, W.J., Mander, Ü., Lu, J.J., Batson, J., Zhang, L., and He, W.S.: Methane
810 emissions from freshwater riverine wetlands, *Ecol. Eng.*, 37:16–24, 2011.

811 Shamir, E., Thompson, T.L, Karpisak, M.M., Freitas, R.J., and Zauderer, J.: Nitrogen
812 accumulation in a constructed wetland for dairy wastewater treatment, *J. Amer. Water*
813 *Resour. Assoc.*, 37(2), 315-325, 2001.

814 Song, K., Lee, S.H., and Kang, H.: Denitrification rates and community structure of denitrifying
815 bacteria in newly constructed wetland, *Eur. J. Soil Biol.*, 47(1), 24-29, 2011.

816 Song, K., Lee, S.H., Mitsch, W.J., and Kang, H.: Different responses of denitrification rates and
817 denitrifying bacterial communities to hydrological pulses in created wetlands, *Soil Biol.*
818 *Biochem.*, 42, 1721–1727, 2010.

819 Søvik, A.K., Augustin, J., Heikkinen, K., Huttunen, J.T., Necki, J.M., Karjalainen, S.M., Klove,
820 B., Liikanen, A., Mander, U., Puustinen, M., Teiter, S., and Wachniew, P.: Emission of
821 the greenhouse gases nitrous oxide and methane from constructed wetlands in Europe. *J.*
822 *Environ. Qual.*, 35, 2360-2373, 2006.

823 Stadmark, J., and Leonardson, L.: Emissions of greenhouse gases from ponds constructed for
824 nitrogen removal, *Ecol. Eng.*, 25, 542-551, 2005.

825 Ström, L., Ekberg, A., Mastepanov M., and Christensen, T.R.: The effect of vascular plants on
826 carbon turnover and methane emissions from a tundra wetland, *Global Change Biol.* 9,
827 1185–1192, 2003.

828 Ström, L., Lampa, A., and Christensen, T.R.: Greenhouse gas emissions from a constructed
829 wetland in southern Sweden, *Wetlands Ecol. Manage.*, 15, 43–50, 2007.

830 Ström, L., Mastepanov, M., and Christensen, T.R.: Species specific effects of vascular plants on
831 carbon turnover and methane emissions from wetlands, *Biogeochem.*, 75, 65–82, 2005.

832 Tai, P.D., Li, P.J., Sun, T.H., He, Y.W., Zhou, Q.X., Gong, Z.Q., Mizuochi, M., and Inamori, Y.:
833 Greenhouse gas emissions from a constructed wetland for municipal sewage treatment, *J.*
834 *Environ. Sci. China*, 14(1), 27–33, 2002.

835 Takaya, N.: Dissimilatory nitrate reduction metabolisms and their control in fungi, *J. Biosci.*
836 *Bioeng.*, 94, 506–510, 2002.

- 837 Tanner, C.C., and Kadlec, R.H.: Influence of hydrological regime on wetland attenuation of
838 diffuse agricultural nitrate losses, *Ecol. Eng.*, 56, 79-88, 2013.
- 839 Tanner, C.C., and Sukias, J.P.S.: Multi-year nutrient removal performance of three constructed
840 wetlands intercepting drainage flows from intensively grazed pastures, *J. Environ. Qual.*,
841 40, 620-633, 2011.
- 842 Tanner, C.C., Nguyen, M.L., and Sukias, J.P.S.: Nutrient removal by a constructed wetland
843 treating subsurface drainage from grazed dairy pasture, *Agric. Ecosyst. Environ.*, 105,
844 145-162, 2005.
- 845 Teiter, S., and Mander, Ü.: Emission of N₂O, N₂, CH₄ and CO₂ from constructed wetlands for
846 wastewater treatment and from riparian buffer zones, *Ecol. Eng.*, 25(5), 528–541, 2005.
- 847 Thayalakumaran, T., Bristow, K.L., Charlesworth, P.B., and Fass, T.: Geochemical conditions in
848 groundwater systems: Implications for the attenuation of agricultural nitrate, *Agril. Water
849 Manage.*, 95, 103-115, 2008.
- 850 Toet, S., Richards, S.P., van Logtestijn, Kamp, R., Schreijer, M., and Verhoeven, J.T.A.: The
851 effect of hydraulic retention time on the performance of pollutants from sewage treatment
852 plant effluent in a surface flow constructed wetland system, *Wetlands*, 25, 375-391, 2005.
- 853 Van der Zaag, A.C., Gordon, R.J., Burton, D.L., Jamieson, R.C., and Stratton, G.W.: Greenhouse
854 gas emissions from surface flow and subsurface flow constructed wetlands treating dairy
855 wastewater, *J. Environ. Qual.*, 39, 460-471, 2010.
- 856 Van Oostrom, A.J, and Russell, J.M.: Denitrification in constructed wastewater wetlands
857 receiving high concentrations of nitrate, *Water Sci. Technol.*, 29, 7-14, 1994.
- 858 Verhoeven, J.T.A., and Meuleman, A.F.M.: Wetlands for wastewater treatment: Opportunities
859 and limitations, *Ecol. Eng.*, 12, 5–12, 1999.
- 860 Vymazal, J., and Kröpfelova, L.: Growth of *Phragmites australis* and *Phalaris arundinacea* in
861 constructed wetlands for wastewater treatment in the Czech Republic, *Ecol. Eng.*, 25,
862 606-621, 2005.
- 863 Vymazal, J., and Kröpfelová, L.: Types of constructed wetlands for wastewater treatment in:
864 Wastewater Treatment in Constructed Wetlands with Horizontal Sub-Surface Flow,
865 Environmental Pollution Springer, Heidelberg, 121-202, 2010.
- 866 Vymazal, J.: Removal of nutrients in various types of constructed wetlands, *Sci. Total Environ.*,
867 380, 48-65, 2007.
- 868 Wang, H., Huang, C., Ge, Y., zhi Wu, J., and Chang, J.: The Performance of Species Mixtures in
869 Nitrogen and Phosphorus Removal at Different Hydraulic Retention Times, *Pol. J.
870 Environ. Stud.*, 23, 917-922, 2014.

871 Weerakoon, G.M.P.R., Jinadasa, K.B.S.N., Herath, G.B.B., Mowjood, M.I.M., and vanBruggen,
872 J.J.A.: Impact of the hydraulic loading rate on pollutants removal in tropical horizontal
873 subsurface flow constructed wetlands, *Ecol. Eng.*, 61, 154-160, 2013.

874 Wild, U., Lenz, A., Kamp, T., Heinz, S., and Pfadenhauer, J.: Vegetation development, nutrient
875 removal and trace gas fluxes in constructed *Typha* wetlands, In: Mander Ü, Jenssen PD,
876 (eds.). *Natural wetlands for wastewater treatment in cold climates*, *Adv. Ecol. Sci.*,
877 Southampton, Boston, WIT Press, p. 101–26, 2002.

878 Wunderlin, P., Lehmann, M.F., Siegrist, H., Tuzson, B., Joss, A., Emmenegger, L., Mohn, J.:
879 Isotope signatures of N₂O in a mixed microbial population system: constraints on N₂O
880 producing pathways in wastewater treatment. *Environ. Sci. Technol.*, 47, 1339–1348,
881 2013.

882 Xue, Y., Kovacic, D.A., David, M.B., Gentry, L.E., Mulvaney, R.L., and Lindau, C.W.: *In situ*
883 measurements of denitrification in constructed wetlands, *J. Environ. Qual.*, 28(1), 263–
884 269, 1999.

885 Yan, C., Zhang, H., Li, B., Wang, D., Zhao, Y., and Zheng, Z.: Effects of influent C/N ratios on
886 CO₂ and CH₄ emissions from vertical subsurface flow constructed wetlands treating
887 synthetic municipal wastewater, *J. Hazardous Materials*, 203-204, 188-194, 2012.

888 Yavitt, J.B., and Knapp, A.K.: Aspects of methane flow from sediment through emergent cattail
889 (*Typha latifolia*) plants, *New Phytol.*, 139, 495-503, 1998.

890 Yin, S., Shen, Q., Tang, Y., and Cheng, L.: Reduction of nitrate to ammonium in selected paddy
891 soils in China, *Pedosphere*, 8, 221–228, 1998.

892 Zhang, J., Shao, W.S., He, M., Hu, H.Y., and Gao, B.Y.: Treatment performance and
893 enhancement of subsurface constructed wetland treating polluted river water in winter,
894 *Environ. Sci. (in Chinese)*, 27, 1560–1564, 2006.

895 Zhao, Y., Zhang, Y., Ge, Z., Hu, C., and Zhang, H.: Effects of influent C/N ratios on wastewater
896 nutrient removal and simultaneous greenhouse gas emission from the combinations of
897 vertical subsurface flow constructed wetlands and earthworm eco-filters for treating
898 synthetic wastewater, *Environ. Sci. Proc. Impacts*, 16, 567-575, 2014.

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Table 1 Mineral N (mg N L⁻¹), total nitrogen (TN, mg N L⁻¹) and TN removal (%) in various CWs treating wastewater; average (\pm standard error, SE) are presented for TN removal; n.a – data not available

CW type	Treatment	N input (mg N L ⁻¹)			N output (mg N L ⁻¹)			N removal (%)			References
		TN	NH ₄ ⁺	NO ₃ ⁻	TN	NH ₄ ⁺	NO ₃ ⁻	TN	NH ₄ ⁺	NO ₃ ⁻	
SF_Finland	Municipal	1.4 \pm 150	0.03 \pm 5.8	0.3 \pm 95	1.1 \pm 48	0.01 \pm 3.0	0.02 \pm 6.7	21.4	66.7	93.3	Søvik et al., 2006
SF_Finland	Agril. runoff	66.1 \pm 1.9	63.5 \pm 1.3	0.7 \pm 0.13	64.7 \pm 1.7	61.2 \pm 1.7	0.3 \pm 0.09	2.1	3.6	57.1	Søvik et al., 2006
SF_Norway	Municipal	43.4 \pm 3.6	41.5 \pm 3.0	0.0 \pm 0.0	36.7 \pm 2.7	32.6 \pm 1.9	0.9 \pm 0.4	15.4	21.4	-800	Søvik et al., 2006
SF	Municipal	n.a	4.5	15.5	n.a	n.a	n.a	61	n.a	n.a	Song et al., 2011
SF	Domestic	n.a	40	5	n.a	n.a	n.a	97-98	n.a	n.a	Dzakpasu et al., 2011
SF	Various	n.a	39	4.4	n.a	n.a	n.a	39-48	n.a	n.a	Vymazal, 2007
SF	Municipal	n.a	36		n.a	n.a	n.a	39	n.a	n.a	Vymazal, 2010
SF	Municipal	n.a	196	<2	n.a	n.a	n.a	35	n.a	n.a	Shamir et al., 2001
SF	various	n.a	80	<1	n.a	n.a	n.a	>90	n.a	n.a	Harrington et al., 2007
SF	Municipal	n.a	0.95	1.54	n.a	n.a	n.a	45	n.a	n.a	Toet et al., 2005
SF	Dairy washout	227	n.a	n.a	n.a	n.a	n.a	40	n.a	n.a	Van der Zaag et al., 2010
All SF								31.3 \pm 6.3			
HSSF_Estonia	Municipal	96.5 \pm 3.0	83.9 \pm 2.7	0.2 \pm 0.02	46.2 \pm 1.5	36.2 \pm 1.4	5.9 \pm 0.65	52.1	56.9	-2850	Søvik et al., 2006
HSSF_Norway	Municipal	53.4 \pm 4.3	38.4 \pm 7.7	14.1 \pm 7.5	45.0 \pm 4.1	43.1 \pm 4.7	1.0 \pm 0.8	15.7	-12.2	92.9	Søvik et al., 2006
HSSF	Dairy washout	306 \pm 101*	n.a	n.a	177 \pm 58*	n.a	n.a	42.2	n.a	n.a	Van der Zaag et al., 2010
HSSF	Domestic	n.a	74.9	3.9	n.a	n.a	n.a	29	n.a	n.a	O’Luanaigh et al., 2010
HSSF	Domestic	87						46-48			Mander et al., 2008
HSSF	Dairy washout	227						28			Van der Zaag et al., 2010
HSSF	Milk parlour	112	22	n.a	24	11	n.a	78	50	n.a	Kato et al., 2006
HSSF	Agriculture	67	40	0.85	27	11	1.1	47	39	-29	Vymazal and Kröpfelova, 2010
HSSF	Industry	124	65	8.5	103	31	7.4	20	20	8	Vymazal and Kröpfelova, 2010
HSSF	Landfill	157	149	1.5	147	98	1.3	30	33	31	Vymazal and Kröpfelova, 2010
HSSF	Municipal	43	24	2	24	14	1.2	40	30	33	Vymazal and Kröpfelova, 2010
All HSSF								40.4 \pm 4.4			
VSSF_Estonia	Municipal	50.9 \pm 9.2	35.7 \pm 6.2	1.1 \pm 0.32	43.1 \pm 7.6	31.7 \pm 5.5	1.7 \pm 0.84	15.3	11.2	-54.5	Søvik et al., 2006
VSSF_Norway	Municipal	52.6 \pm 5.2	49.6 \pm 4.0	0.0 \pm 0.0	47.8 \pm 6.9	21.4 \pm 6.9	25.5 \pm 1.3	9.1	56.9	-25400	Søvik et al., 2006
VSSF	Municipal	41.0 \pm 0.5	n.a	n.a	20.7 \pm 0.8	n.a	n.a	49.3 \pm 1.8	n.a	n.a	Yan et al., 2012
VSSF	Municipal	46 \pm 13	n.a	n.a	n.a	n.a	n.a	74 \pm 3	n.a	n.a	Zhao et al., 2014
All VSSF								37.0 \pm 10.9			

SF- surface flow; HSSF – horizontal subsurface flow; VSSF- vertical subsurface flow; *mg N m⁻² h⁻¹

Table 2 Total organic C (TOC) removal (%) in various CWs treating wastewater; average (\pm standard error, SE) are presented for TOC removal; n.a - data not available

CWs type	Treatment	C input (TOC; mg C L ⁻¹)	C outflow (TOC; mg C L ⁻¹)	TOC Removal (%)	References
SF_Finland	Municipal	13.0 \pm 0.3	14.0 \pm 0.5	-7.7	Søvik et al., 2006
SF_Finland	Agril runoff	25.0 \pm 3.4	20.0 \pm 3.4	20.0	Søvik et al., 2006
SF_Norway	Municipal	26.7 \pm 2.9	17.1 \pm 1.8	36.0	Søvik et al., 2006
SF	Dairy wash out	186 [†]	136 [†]	27	Van der Zaag et al., 2010
All SF				18.8 \pm 9.4	
HSSF	Domestic	150*	n.a	n.a	Garcia et al., 2007
HSSF	Dairy wash out	186 [†]	107.9 [†]	42	Van der Zaag et al., 2010
HSSF_Estonia	Municipal	62.8 \pm 16.6 [†]	41.0 \pm 11.3 [†]	34.7	Søvik et al., 2006
HSSF_Norway	Municipal	40.5 \pm 11.3	15.0 \pm 2.4	63.0	Søvik et al., 2006
All HSSF				46.6 \pm 7.3	
VSSF_Estonia	Municipal	132.2 \pm 32.2 [†]	62.8 \pm 16.6 [†]	52.5	Søvik et al., 2006
VSSF_Norway	Municipal	40.5 \pm 11.3	15.0 \pm 2.4	63.0	Søvik et al., 2006
VSSF	Municipal	106 \pm 35	74 \pm 21	26 \pm 4.6	Yan et al., 2012
VSSF	Municipal	249 \pm 49	n.a	83 \pm 1.0	Zhao et al., 2014
All VSSF				56.2 \pm 9.5	

SF- surface flow; HSSF – horizontal subsurface flow; VSSF- vertical subsurface flow; [†]BOD; *mg m⁻² h⁻¹

Table 3 Nitrous oxide (N₂O) emissions (mg N m⁻² d⁻¹); N₂ emissions (mg N m⁻² d⁻¹) and N₂O emission factor (N₂O/TN input*100) in various type of CWs; mean (± SE) was presented for N₂O emission factor; n.a - data not available

CW type	Treatment	Denitrification		N ₂ O-N/TN (%)	N ₂ -N/TN (%)	References
		N ₂ O emissions (mg N m ⁻² d ⁻¹)	N ₂ emissions (mg N m ⁻² d ⁻¹)			
HSF	Agril. tile drainage	0.01-0.12	n.a	0.19-1.4	n.a	Xue et al., 1999
HSF	Treated municipal	2.0±3.3	n.a	0.02-0.27	n.a	Johansson et al., 2003
HSF	Agril. drainage	-0.2-1.9	n.a	-0.14-0.52	n.a	Wild et al., 2002
HSF	Dairy wash out	16.8±7.0	n.a	0.33±0.12	n.a	Van der Zaag et al., 2010
HSF_Finland	Municipal	0.01±0.01	n.a	1.6±1.3	n.a	Søvik et al., 2006
HSF_Finland	Agril. runoff	0.40±0.25	n.a	0.37±0.18	n.a	Søvik et al., 2006
HSF_Norway	Municipal	4.0±1.6	n.a	1.5±4.4	n.a	Søvik et al., 2006
All SF		2.78±1.72		0.61±0.21		
HSSF	Domestic	0.2-17.0	n.a	0.06-3.8	n.a	Mander et al., 2005
HSSF_Estonia	Municipal	7.1±1.2	n.a	0.05±0.31	n.a	Søvik et al., 2006
HSSF_Norway	Municipal	6.9±4.3	n.a	0.24±0.53	n.a	Søvik et al., 2006
HSSF	Domestic	1.3-1.4	160-170	0.37-0.60	15.2-22.7	Mander et al., 2008
HSSF	Domestic	0.003-0.001	0.01-5.42	n.a	n.a	Teiter and Mander, 2005
HSSF	Domestic	0.13	n.a	0.008	n.a	Fey et al., 1999
HSSF	Dairy wash out	9.5±1.5	n.a	0.18±0.12	n.a	Van der Zaag et al., 2010
HSSF	Domestic	0.17	n.a	0.23	n.a	Liu et al., 2009
VSSF	Domestic	0.17	n.a	0.01		Mander et al., 2011
All HSSF		4.23±1.87		0.62±0.38		
VSSF	Domestic	0.001-0.002	0.01-5.0	n.a	n.a	Teiter and Mander, 2005
VSSF	Domestic	4.6	150	0.45-0.50	n.a	Mander et al., 2008
VSSF	Domestic	11.0	n.a	0.29	n.a	Mander et al., 2005
VSSF	Domestic	1.44	n.a	0.03		Mander et al., 2011
VSSF	Domestic	0.005	n.a	0.09	n.a	Gui et al., 2007
VSSF	Domestic	0.003	n.a	0.04	n.a	Liu et al., 2009
VSSF_Estonia	Municipal	15±3.9	n.a	04.3±0.95	n.a	Søvik et al., 2006
VSSF_Norway	Municipal	960±40	n.a	1.4±0.72	n.a	Søvik et al., 2006
All VSSF		123.8±106		1.01±0.48		

SF- surface flow; HSSF – horizontal subsurface flow; VSSF- vertical subsurface flow

Table 4 Carbon dioxide (CO₂, mg C m⁻² d⁻¹), CH₄ (mg C m⁻² d⁻¹) and CH₄ emission factor (CH₄-C/TOC input*100) in various type of CWs; mean (± SE) was presented for CH₄ emission factor; n.a - data not available

CWs type	Treatment	CO ₂ emissions (mg C m ⁻² d ⁻¹)	CH ₄ emissions (mg C m ⁻² d ⁻¹)	CH ₄ / TC (%)	References
SF	Municipal	n.a	5.4	n.a	Tai et al., 2002
SF	Domestic	0.19	n.a	26	Gui et al., 2007
SF	Domestic	1.13	n.a	16	Liu et al., 2009
SF	Agril. drainage	n.a	0.88	31	Wild et al., 2002
SF	Dairy wash out	4250±550	223±35	9.45	Van der Zaag et al., 2010
SF_Finland	Municipal	1200±420	29±6.4	19±4.3	Søvik et al., 2006
SF_Finland	Agril runoff	3200±560	350±180	11±5.5	Søvik et al., 2006
SF_Norway	Municipal	1400±250	72±28	4.8±2.2	Søvik et al., 2006
All SF		1675±703	113±58	16.8±3.8	
HSSF	Domestic	n.a	1.7-528	n.a	Mander et al., 2005a
HSSF	Domestic	2.54-5.83	0.03-0.40	n.a	Teiter and Mander, 2005
HSSF	Domestic	5.33	0.001	0.03	Garcia et al., 2007
HSSF	Domestic	n.a	0.03	4.3	Gui et al., 2007
HSSF	Domestic	n.a	0.29	4.0	Liu et al., 2009
HSSF	Dairy wash out	3475±375	118±9.0	4.4	Van der Zaag et al., 2010
HSSF	Domestic	0.6-1.7	1.4- 4.1	0.12-0.23	Søvik et al., 2006
HSSF	Domestic	600	0.48	0.02	Mander et al., 2011
HSSF_Estonia	Municipal	3800±210	340±240	n.a	Søvik et al., 2006
HSSF_Norway	Municipal	790±170	130±43	9.5±3.3	Søvik et al., 2006
All HSSF		1010±672	112±74	3.23±1.4	
VSSF	Domestic	5.83-12.13	0.60-5.70		Teiter and Mander, 2005
VSSF	Domestic	n.a	16.4	n.a	Mander et al., 2005
VSSF	Domestic	n.a	0.013	1.68	Gui et al., 2007
VSSF	Domestic	n.a	0.13	1.73	Liu et al., 2009
VSSF	Municipal	2662 ± 175	33.5 ± 3.2	n.a	Mander et al., 2008
VSSF	Domestic	1080	3.36	0.05	Mander et al., 2011
VSSF_Estonia	Municipal	8400±2100	110±35	n.a	Søvik et al., 2006
VSSF_Norway	Municipal	22000±5000	140±160	0.39±0.27	Søvik et al., 2006
All VSSF		6616 ± 3779	42.9 ± 23.7	1.27±0.31	

SF- surface flow; HSSF – horizontal subsurface flow; VSSF- vertical subsurface flow

Table 5 Nitrous oxide (N₂O, mg N m⁻² d⁻¹), CO₂ and CH₄ emissions (mg C m⁻² d⁻¹) in various type of CWs under different plant types; n.a - data not available

CW type	Wastewater type	Plant type	N ₂ O (mg N m ⁻² d ⁻¹)	CH ₄ (mg C m ⁻² d ⁻¹)	CO ₂ (mg m ⁻² d ⁻¹)	Reference
HSF	Secondary treated municipal	No plant	3.79±2.64	163±209		Johansson et al., 2003; Johansson et al., 2004
		<i>Typha lotifolia</i>	2.64±4.09	109±185	n.a	
		<i>Phalaris arundinacea</i>	3.79±3.44	212±151	n.a	
		<i>Glyceria maxima</i>	0.76±1.01	112±178	n.a	
		<i>Lemna minor</i>	1.45±1.18	450±182	n.a	
		<i>Spirogyra sp.</i>	0.98±1.25	107±135	n.a	
HSF	Sewage Treatment water	No plant	-0.26±2.53	-4.76±61.8	4.32±0.73	Ström et al., 2007
		<i>Typha atifolia</i>	4.94±2.00	225±47.7	25.3±4.08	
		<i>Phragmites australis</i>	7.80±2.53	333±76.6	25.1±4.74	
		<i>Juncus effusus</i>	3.87±1.86	489±46.3	26.1±3.00	
HSSF	Domestic	No plant	0.04±0.02	87±6.3	80±6.3	Maltais-Landry et al., 2009
		<i>Phragmites</i>	0.06±0.03	50±7.5	200±35	
		<i>Typha</i>	0.03±0.01	28±3.0	235±32	
		<i>Phalaris</i>	0.01±0.01	45±6.0	195±31	
VSSF	Municipal	<i>Phragmites australis</i>	15±3.9	110±35	8400±2100	Søvik et al., 2006
VSSF	Municipal	<i>Phragmites australis</i>	264	384		Mander et al., 2005

SF- surface flow; HSSF – horizontal subsurface flow; VSSF- vertical subsurface flow

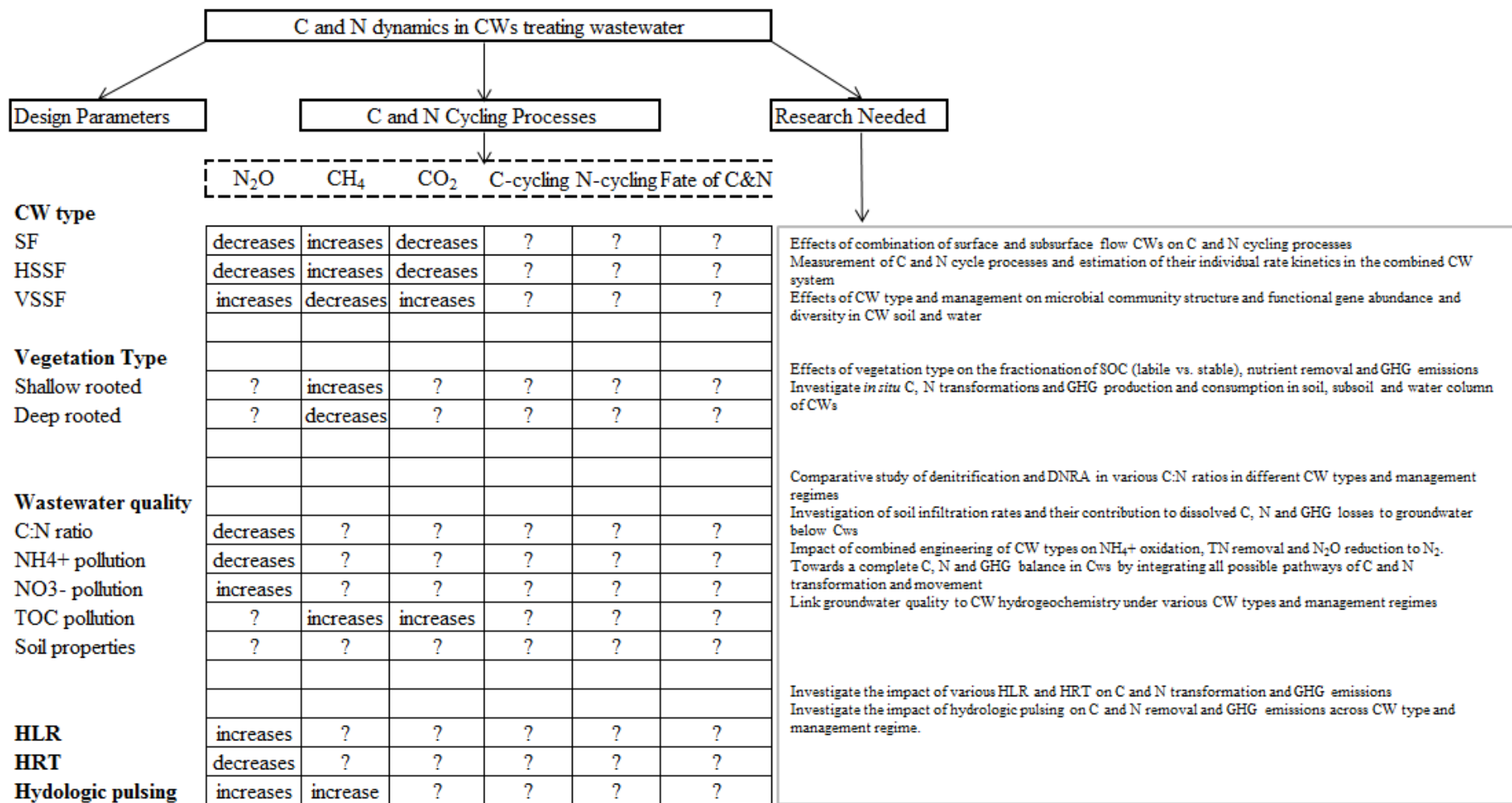


Figure 1 Conceptual model showing the current state of knowledge of C and N dynamics in constructed wetlands treating wastewater and the specific experimental work that needs to be undertaken in the future; SF- surface flow; HSSF – horizontal subsurface flow; VSSF- vertical subsurface flow; HLR – hydraulic loading rate; HTR – hydraulic retention time; ? – not known or very little known

Blackburn, T. H. (1979). "Methods for measuring rates of NH_4^+ turnover in anoxic marine sediments, using a $^{15}\text{N-NH}_4^+$ dilution technique." Applied and Environmental Microbiology 37(4): 760-765.

Huygens, D., M. Trimmer, et al. (2013). Biogeochemical N cycling in wetland ecosystems: ^{15}N isotope techniques. Methods in biogeochemistry of wetlands. K. R. Reddy, J. P. Megonigal and R. D. Delaune, Soil Science Society of America. 30: 553-591.

Müller, C., R. J. Laughlin, et al. (2014). A ^{15}N tracing method to quantify N_2O pathways from terrestrial ecosystems. European Geological Union General Assembly. EGU. Vienna, European Geological Union: 1.

Rütting, T. and C. Müller (2008). "Process-specific analysis of nitrite dynamics in a permanent grassland soil by using a Monte Carlo sampling technique." European Journal of Soil Science 59: 208-215.